

THE THERAPEUTIC POTENTIAL OF BRADYKININ B₁ RECEPTOR ANTAGONISTS IN CHRONIC PAIN

S. Farkas and J. Éles

Research Division, Gedeon Richter, Plc, Budapest, Hungary

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SUMMARY

After the rise and fall of cyclooxygenase-2 (COX-2)-selective inhibitors as safe replacements for nonsteroidal antiinflammatory drugs (NSAIDs), particularly for the treatment of chronic painful conditions in the elderly, there is still a great need for safer drugs that can alleviate both inflammation and pain. Des-arginine metabolites of bradykinin-related peptides are involved in the propagation and maintenance of inflammation and pain via activation of bradykinin B₁ receptors, the bradykinin receptor subtype that is hardly detectable in healthy tissues but induced during inflammation. Due to the really inducible nature of the receptor at peripheral sites, orally acting B₁ receptor antagonists hold the promise of being a safe substitute for NSAIDs, especially for chronic treatment. In addition, nonclinical research suggests the potential utility of central nervous system-penetrating B₁ receptor antagonists in a broader range of chronic pain states, including neuropathic pain, and involving a central sensitization process. However, exploitation of the therapeutic utility has been hindered by the extreme difficulties related to optimization for an inducible peptide-receptor with species-specific differences in physiology and pharmacology. The scientific literature implicating therapeutic utility for B₁ receptor antagonists and the ongoing efforts for exploiting this target are reviewed in this article.

INTRODUCTION

Bradykinin and related peptides, kinins, have long been acknowledged as algogenic mediators released at sites of inflammation and/or tissue injury. The inflammatory response functions as a two-edged sword in several pathophysiological processes. Namely, it serves the beneficial purpose of defense and repair, either by eliciting pain and consequent avoidance of further injury, or by promoting cellular responses serving immunological defense against intruders, like infections or tumor cells, as well as by promoting repair and healing of the damaged tissues. However, this homeostatic response can go wrong in many pathological situations, such that pain, inflammation and/or an overly active immune response becomes a chronic condition, promoting maintenance of a disease state instead of healing. Therefore, it is a great challenge for pharmacotherapy to effectively mitigate chronic pain and inflammation without impairing the defense and repair functions. Inflammation is a very complex programmed process, in which multiple humoral mediators play in concert to initiate, propagate, maintain and resolve this state and its various phases. The kallikrein-kinin system is one of the multiple systems that play a role in the initiation, propagation and maintenance of inflammation and pain, and as such, may provide a promising target for pharmacological intervention (1).

THE KALLIKREIN-KININ SYSTEM

The kallikrein-kinin system is part of a humoral defense system and participates in the inflammatory response in various organs. It represents a proteolytic complex that, when activated, triggers the release of kinins. In mammals, endogenous kinins include the nonapeptide bradykinin (BK; Arg-Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg), the decapeptide lysyl-bradykinin (LysBK; Lys-Arg-Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg; also termed kallidin) and their respective carboxy-terminal (right end) des-arginine metabolites: des-arginine-bradykinin (DABK) and lysyl-des-arginine-bradykinin (LysDABK). There are two types of kallikreins: plasma and tissue kallikreins. Both are serine protease enzymes that cleave precursor proteins, "kininogens", to release kinins. The plasma kallikrein system is involved in triggering the intrinsic coagulation cascade and consists of three serine proenzymes (factors FXII, FXI and prekallikrein) and high-molecular-weight kininogen (HMWK). Contact between plasma and a negatively charged surface, which occurs at sites of blood vessel injury, induces the activation of factor XII to factor XIIa, subsequently stimulating the conversion of prekallikrein to kallikrein, followed by the

cleavage of HMWK to release BK, in addition to triggering the coagulation cascade. Another mechanism for initiation of the activation of the kallikrein–kinin system depends on binding of components of the contact activation cascade on the surface of cells, such as leukocytes, platelets, endothelial cells and myocytes (1, 2).

The tissue kallikrein system is widely distributed throughout the body. Tissue kallikrein preferentially cleaves low-molecular-weight kininogen to generate LysBK. The mechanisms that regulate the tissue kallikrein activity are poorly understood (1). Regulation mechanisms include secretion and codistribution of a kallikrein inhibitor protein, kallistatin (3, 4). However, abundant evidence indicates that the kallikrein–kinin system is activated in response to tissue injury, infection and inflammation, leading to local release of kinins.

BRADYKININ RECEPTORS

The released kinins exert various effects via two G protein-coupled receptors –the bradykinin B₁ and B₂ receptors (5, 6)–, the existence of which has been confirmed by expression cloning experiments in several species, including mice (7, 8), rats (9, 10), rabbits (11, 12), dogs (13) and humans (14–16). Although the existence of a third bradykinin receptor (B₃) had been postulated (17), it could not be validated by cloning and was finally refuted (18). The released primary kinin peptides, BK and LysBK, are metabolized by cleavage at several sites (19). However, two metabolic pathways are dominant. Primary kinins are either metabolized to inactive peptides by angiotensin-converting enzyme (ACE, kininase II) or converted by carboxypeptidase enzymes (kininase I) via removal of the carboxy-terminal arginine residue to the des-arginine metabolites (20), DABK and LysDABK, which are also biologically active kinins with altered affinities for the bradykinin receptor subtypes. Affinities of the primary kinins and their des-arginine metabolites for the two subtypes of bradykinin receptors cloned from different species, as well as their functional agonist potencies at the human receptors, are summarized in Table I.

In principle, the primary kinins are potent and selective agonists for B₂ receptors, but have low affinity for the B₁ receptor. On the contrary, the des-arginine products are potent agonists of the B₁ receptor, but have mostly low affinity for B₂ receptors. In particular, the affinity of BK is so low for B₁ receptors that these receptors are in fact not bradykinin but “only” kinin receptors. It is worth noting here that estimated BK + LysBK and DABK + LysDABK tissue concentrations in inflamed rat (27) or human (28) tissues, or in rabbit plasma, after treatment with bacterial lipopolysaccharides (29) range at approximately 0.05–2 nmol/kg (or nmol/L), and hence, hardly exceed the low nanomolar range. Formerly, plasma and tissue concentrations of “BK-related material” were assessed by radioimmunoassay or bioassay in biological fluids from healthy or diseased patients/volunteers and were found to range from subnanomolar to 55 nM (58 ng/mL) (6), the latter being observed in synovial fluid from rheumatoid arthritis (RA) patients during exacerbation (30). Interestingly, although DABK has equal or higher affinity than LysDABK for rodent (rat and mouse) and dog B₁ receptors, it has two orders of magnitude lower affinity in rabbits and negligible relative affinity in humans. Despite this, DABK appeared as a potent functional B₁ receptor agonist at cloned recombinant human B₁ receptors. However, in functional studies of native B₁ receptors in human umbilical vein endothelial cells, DABK had only moderate potency, two orders of

Table I. Affinity and functional potency values of natural kinin peptides in various species.

	Species	Affinities (K _i , nM)			
		BK	LysBK	DABK	LysDABK
B ₁	Mouse (8, 21)	> 1,000	510	0.3–0.7	1.3–1.7
	Rat (10)	120	–	2.0	2.4
	Rabbit (12)	> 5,000	19	32	0.23
	Dog (13)	520	440	4.7	1.6
	Rhesus (22)	2,200	110	350	0.17
	Human (23)	> 10,000	1.35	1370	0.12
B ₂	Mouse (7)	0.48	0.52	8.1	6,400
	Rat (24)	0.07	0.09	> 100,000	–
	Rabbit (11)	≈ 3	≈ 1.5	> 1,000	> 1,000
	Dog (13)	3.9	2.1	> 10,000	> 10,000
				8,100	
	Human (7, 24)	0.12–0.54	0.30–0.63	69,000	> 30,000
Functional potencies at recombinant receptors (EC ₅₀ , nM)					
		BK	LysBK	DABK	LysDABK
B ₁	Human (25)	1,000	243	8.6	7.9
B ₂	Human (25)	2	10	> 3,000	> 10,000
Functional potencies on human umbilical vein (EC ₅₀ , nM)					
		BK	LysBK	DABK	LysDABK
B ₁	Human (26)	> 5,000	> 5,000	204.2	2.5
B ₂	Human (26)	2.5	1.5	> 5,000	> 5,000

BK, bradykinin; LysBK, lysyl-bradykinin; DABK, des-arginine-bradykinin; LysDABK, lysyl-des-arginine-bradykinin.

magnitude less than LysDABK (see Table I). On the other hand, LysBK appeared to have rather high affinity but low functional potency at human B₁ receptors. In brief, both BK and LysBK are potent and selective B₂ receptor agonists, whereas LysDABK is a potent and selective B₁ receptor agonist in all species. DABK is also a potent and selective B₁ receptor agonist in mice, rats and dogs, but not in rabbits, monkeys and humans. The B₁ receptor-stimulating des-arginine products are also inactivated, mainly by ACE. However, while the natural B₂ receptor agonists have short half-lives (e.g., 10–50 s for BK), the natural B₁ receptor agonists have 4- to 12-fold longer half-lives (31, 32). As a consequence, in circumstances associated with chronic activation of the kallikrein–kinin system the B₁ receptor agonist peptides are more prone to accumulation.

The regulation of the expression and activity of B₁ and B₂ receptors is remarkably different. Bradykinin B₂ receptors are constitutively expressed in a wide variety of tissues and cell types. Moreover, B₂ receptors, when exposed to bradykinin, undergo rapid desensitization and internalization (19). Therefore, they are more predestined for mediating acute-phase responses immediately following injury and initiation of inflammation. In contrast, B₁ receptors are expressed at a very low level, if at all, in healthy tissues, but undergo massive upregulation (or if they lack constitutive expression, “induction”) upon tissue injury or inflammation. Nevertheless, B₁ receptors are not prone to desensitization or agonist-induced recep-

tor internalization. These features, i.e., activation by secondary kinins, which have a longer half-life, and the inducible character, predestine B₁ receptors more for mediating subacute and chronic phases of inflammation and associated pain. However, it is worth noting that there are some exceptions to this general rule. The constitutive presence of B₁ receptors in blood vessels of dogs (33-35) and in the sensory neurons, spinal cord and other central nervous system (CNS) sites of rats, primates and humans has also been demonstrated (36-41). Moreover, upregulation of B₂ receptors has also been described under some conditions, for example in neuropathies (41, 42), simultaneously with B₁ receptor induction. However, desensitization and internalization remain limiting factors of long-term effects via B₂ receptors, particularly in inflammation.

Several proinflammatory/injury mediators have a documented role in induction of B₁ receptors, including growth factors, such as epidermal growth factor (EGF) (43) and glial cell line-derived neurotrophic factor (GDNF) (44), interferon- γ (45) and cytokines, including TNF- α , IL-1 β , IL-6 and IL-8 (46-48). Moreover, B₁ receptors are autoinduced by B₁ and B₂ kinin agonists. Furthermore, IL-1 β and LysDABK can act synergistically in a supra-additive fashion to boost B₁ receptor induction (49), and priming with bradykinin also leads to B₁ receptor induction in vivo (50), as well as in vitro (46, 51), and to evolving function of B₁ receptors. The induction elicited by B₂, but not B₁, receptor agonists appeared to be IL-1 β -dependent (46). The induction mechanisms involve activation of nuclear factor NF- κ B-mediated promoter regulation (52, 53); however, several other mechanisms have also been postulated (54, 55). Topical or systemic exposure to bacterial endotoxins, such as lipopolysaccharide (LPS), is also a strong inducer, mostly via stimulation of cytokine release (56). In turn, kinins stimulate the synthesis and release of nitric oxide (NO), cytokines (57), arachidonic acid, prostaglandins (58), prostacyclin (PGI₂) (59, 60), leukotrienes (59), as well as other chemotactic factors (61).

The presence of both B₁ and B₂ kinin receptors has been documented in diverse tissues and cell types, including endothelial cells (62), vascular (59, 63), gastrointestinal (64, 65) and smooth muscle cells, fibroblasts (46, 66), osteoblasts (67), mucosal epithelium (68, 69), glandular tissues/cells (68, 70) and various blood and inflammatory cells, i.e., neutrophils (71, 72) and eosinophils (73), granulocytes, lymphocytes (45), dendritic cells (74), macrophages (68, 69) and mast cells (75), both in animals and humans. Moreover, expression of both kinin receptors was also shown in the CNS. Constitutive B₁ receptor expression was found in neurons of the thalamus, hypothalamus and spinal cord of rats, monkeys and humans (36, 39, 40, 76, 77). Bradykinin B₁ receptor expression was also found in sensory ganglia (37, 78), and in the spinal cord, B₁ receptor immunoreactivity is confined to spinal dorsal horn neurons and central terminals of afferent fibers (39). Most attempts were unsuccessful at demonstrating a direct functional effect of B₁ receptors on peripheral sensory nerves (79, 80) or nociceptors, even after inducing stimuli. However, recently, potentiation of heat-evoked current by DABK could be demonstrated in GDNF-treated sensory neurons (44). This contrasts with the well-documented constitutive presence and sensitizing function of B₂ receptors on primary afferents (81). Bradykinin B₁ and B₂ receptors are also present in glial cells (82).

The cellular effects of kinin receptor activation include either contraction or relaxation of smooth muscle cells, production and release of

NO and inflammatory mediators, direct or indirect activation and sensitization of sensory nerves (83), and stimulation of cell proliferation or collagen synthesis (84, 85). These effects occur via numerous signaling pathways. At first glance, the major signals generated by the B₁ and B₂ receptors appear to be identical, with the reservation that normally only B₂ receptor-mediated effects can be observed, but a switch occurs to major involvement of B₁ receptors under inflammatory conditions. However, differential signaling pathways for B₁ and B₂ receptors have also been postulated (62). Signaling via activation of phospholipase C (PLC), leading to the formation of diacylglycerol (DAG) and inositol 1,4,5-trisphosphate (IP₃), which increases cytoplasmic Ca²⁺ concentrations, is well defined in numerous cell types (83). DAG activates protein kinase C (PKC), which modulates the function of other proteins, such as the thermo- and capsaicin-sensitive transient receptor potential channel TRPV1 of nociceptors. The increased cytoplasmic Ca²⁺ concentrations participate in the release of various mediators (cytokines, chemokines, NO) or elicit contraction of smooth muscle cells. Release of NO from the endothelium mediates vasodilation of blood vessels. Metabolism of DAG yields products of the arachidonic acid cascade. Kinin agonists also activate phospholipase PLA₂, which releases arachidonic acid, leading to formation of vasodilator prostaglandins such as PGI₂ (83). NO and PGI₂ are thought to be major endothelium-derived vasodilators, but endothelial-derived hyperpolarizing factors also play a role (86). Furthermore, several signaling pathways of kinin receptors involving the extracellular signal-regulated kinase and mitogen-activated protein kinase (ERK/MAPK) and β -arrestin pathways are reviewed and discussed in more detail elsewhere (62, 83, 87).

EXPERIMENTAL TOOLS FOR ELUCIDATING THE ROLES OF B₁ RECEPTORS

Exploration of the role of B₁ receptors in pathophysiology and exploitation of their potential therapeutic utility has been hindered by the difficulty of making good bioavailable pharmacological agents (peptidomimetics) for this peptide receptor and also by the significant species-specific differences in the sequence, structure and pharmacological sensitivity of B₁ receptors. Amino acid sequence identities of B₁ receptors of different species compared to the human sequence are as follows: mouse 68% (21), rat 71% (10), dog 76% (13) and rabbit 78% (12), whereas rhesus and African green monkeys have a very high sequence identity of 97% (22). For comparison, the sequence identity between mouse and rat B₁ receptors is 89% (10), and that between human B₁ and B₂ receptors is only 36% (16). Pharmacological studies with peptide ligands suggested that there are two groups of B₁ receptor subtypes or classes defined by species. One group –“rodent-type”– comprises rat, mouse, hamster and dog, and the other –“human-type”– comprises rabbit, pig, monkey and human (88, 89). Therefore, several classes of new B₁ receptor antagonists designed to act on human B₁ receptors are inactive or weakly active on rodent receptors and thus cannot be used in rats and mice, the most feasible species for nonclinical studies. To circumvent this obstacle, researchers have either used antagonists as pharmacological tools, which are also potent against rodent B₁ receptors, or applied non-rodent animal models using either rabbits, or exceptionally monkeys, for in vivo pharmacological experiments, or developed transgenic animal approaches.

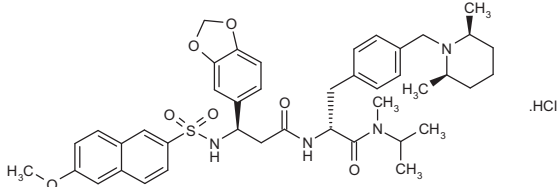
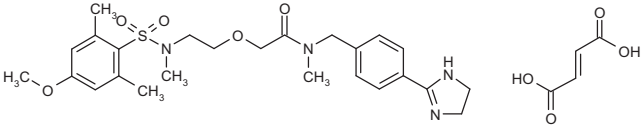
Pharmacological tools

Apart from the use of B₁ receptor knockout mice, the vast majority of studies used peptide or nonpeptide B₁ receptor antagonists to reveal the participation of B₁ receptors in pathophysiological processes. For proper interpretation of the results, it is essential to consider the properties, i.e., virtues and limitations, of these pharmacological tools. The agonists used in exploration of the field were mostly restricted to the natural peptide agonists DABK and LysDABK, although recently, further optimized synthetic peptide agonists were also proposed (90). Table II summarizes the main characteristics of most B₁ receptor antagonist compounds used for *in vivo* studies.

The first potent B₁ receptor antagonists, DALBK and LysDALBK, were obtained by replacing phenylalanine at position 8 with leucine in the

natural B₁ receptor agonists. Although they are pure antagonists at human and rabbit B₁ receptors, in rats and mice they proved to be partial agonists with considerable intrinsic activity (see references in Table II). Some early studies on analgesic effects in rats revealed dose-dependent effects for DALBK, with bell-shaped dose–response curves peaking around 10–30 nmol/kg *s.c.* (102, 103) or 100 nmol/kg *i.p.* (104). This finding may be due to the fact that at low doses it behaves as an antagonist but at higher doses the agonist effect becomes significant (19). On the other hand, much higher doses of DALBK have often been used in various studies with apparent analgesic effects. The interpretation of these effects may be questioned, since low doses of the agonist DABK can also cause analgesia in certain circumstances (105). Therefore, conclusions drawn with pure antagonists may be more reliable. A second gener-

Table II. Selective B₁ receptor antagonists used for elucidation of function *in vivo*.

Name/code	Structure	Characteristics
<i>Peptide, first-generation</i>		
des-Arg9-[Leu8]-BK (DALBK; B6769)	Arg-Pro-Pro-Gly-Phe-Ser-Pro-Leu	Most extensively studied <i>in vivo</i> in rodents. Effective by topical, <i>i.v.</i> , <i>i.p.</i> , <i>s.c.</i> administration. K_i : 3 nM (rat) (10), 4 nM (mouse), 90 nM (rabbit), 130 nM (human) (8). Partial agonist at mouse, rat and dog but pure antagonist at rabbit and human B ₁ receptors (13, 25, 33, 64, 91, 92).
Lys-des-Arg9-[Leu8]-BK (LysDALBK)	Lys-Arg-Pro-Pro-Gly-Phe-Ser-Pro-Leu	Rabbit/human-preferring antagonist. K_i : 7.5 nM (mouse), 0.4 nM (rabbit), 0.4 nM (human) (8). Partial agonist at “rodent-type” but not at “human-type” B ₁ receptors.
<i>Peptide, second-generation</i>		
des-Arg-Hoe 140	[D-Arg]-Arg-Pro-Hyp-Gly-Thi-Ser-[D-Tic]-Oic	More stable than first-generation ligands (93). K_i : 24 nM (mouse), 20 nM (rabbit), 20 nM (human). Weak partial agonist at rat B ₁ receptors (89). Occasionally used <i>in vivo</i> (94).
<i>Peptide, third-generation</i>		
R-715	[Ac-Lys]-Arg-Pro-Pro-Gly-Phe-Ser-[β-D-Nal]-Ile	More stable and potent than first-generation ligands. K_b : 3 nM (rabbit and human), 100 nM (mouse). Pure antagonist! (95–97). Second most widely used peptide B ₁ receptor antagonist <i>in vivo</i> after DALBK. Effective by topical, <i>i.v.</i> , <i>i.p.</i> and <i>s.c.</i> administration. Considered to be not brain penetrant (98).
R-954	[Ac-Orn]-Arg-Oic-Pro-Gly-[αMe-Phe]-Ser-[β-D-Nal]-Ile	Features similar to R-715. Less widely used. More stable against kinase II from platelets (97).
<i>Nonpeptide</i>		
SSR-240612		Potent and selective B ₁ receptor antagonist in all species. K_i : 0.5 nM (human); K_b : 0.4 nM (rat), 1.3 nM (rabbit). Weak B ₂ receptor antagonist, K_i : 358 nM. Orally active in a variety of <i>in vivo</i> models (99). May be brain penetrant to some extent (?) (100). Most widely used nonpeptide B ₁ receptor antagonist.
LF-22-0542		Potent and selective B ₁ receptor antagonist in all species. K_b : 0.6 nM (human), 2 nM (mouse), 3 nM (rat). Not orally active but effective by <i>s.c.</i> and <i>i.p.</i> route (101). Considered to be brain penetrant (98).

ation of B₁ receptor antagonists with somewhat improved metabolic stability is represented by the des-arginine product of the peptidic B₂ receptor antagonist Hoe-140. However, the third-generation peptidic B₁ receptor antagonists R-715 and R-954, which are devoid of partial agonist effects and are metabolically more stable, are better research tools for elucidating functions of B₁ receptors. SSR-240612 was the first orally active nonpeptide B₁ receptor antagonist ligand, which has recently gained popularity as a research tool, as it is almost equally potent on rat and human B₁ receptors (99). Its brain penetration has not been publicly documented, but some pharmacodynamic studies suggest access to central B₁ receptors (100). LF-22-0542 is another nonpeptide B₁ receptor antagonist with high potency on rodent B₁ receptors (101). Although not documented, it is discussed as a brain-penetrating antagonist (98). Nevertheless, it is effective only by the parenteral route of administration. Although LF-22-0542 caused minimal sedation and motor impairment at a high dose (100 mg/kg s.c.), there appears to be sufficient separation from analgesic effects (≥ 10 mg/kg) (101). These nonpeptide antagonists are also devoid of partial agonist activity and their recent use has greatly contributed to exploration of B₁ receptor functions.

Non-rodent in vivo experiments

Many nonpeptide B₁ receptor antagonists with potent activity at the human receptor also have high potency at rabbit but low potency at rat and mouse receptors. For in vivo pharmacodynamic characterization of such compounds, some research groups invented novel preclinical models in rabbits. For example, a rabbit spinal reflex measurement after induction with complete Freund's adjuvant (CFA) was established by Merck scientists (106) and a CFA-induced mechanical hyperalgesia test was applied recently by Amgen scientists. We also published the use of a new rabbit model measuring weight-bearing incapacitation of forelimbs after CFA-induced arthritis (107). Due to the high sequence homology and pharmacological similarity between monkey and human B₁ receptors, a few studies were performed in monkeys. Merck researchers used African green monkeys (108) or rhesus monkeys (109) for in vivo B₁ receptor antagonist potency studies on the hypotensive effects of the B₁ receptor agonist LysDABK after priming with LPS. Anti-hyperalgesic effects were measured in rhesus monkeys in a carrageenan-induced thermal hyperalgesia test by an Elan research group (89).

Transgenic animals

Bradykinin B₁ receptor knockout mice, which were reported to develop in good health but exhibited hypoalgesia and blunted inflammatory responses (110), have been extensively used for exploration of B₁ receptor function by academic groups. For studying the effects of specific human receptor antagonists, "humanized" transgenic mice and rats have been developed. Merck scientists generated a transgenic rat that overexpresses human B₁ receptors in neural tissues (111) for the study of human B₁ receptor occupancy in the rat brain. Unfortunately, these rats were unsuitable for pain studies and attempts to develop a brain-penetrating radioligand have failed. However, this rat model appeared to be suitable for ex vivo occupancy studies (112). For analgesia studies, both Novartis and Merck researchers have developed and used a mouse line in which the mouse B₁ receptor is replaced with the human B₁ receptor and its expression is regulated by the same promoter that is inducible by inflammatory stimuli (113, 114).

PATHOPHYSIOLOGICAL ROLES OF KININ RECEPTORS WITH PARTICULAR RESPECT TO B₁ RECEPTORS

Exogenous injection of BK into animal or human tissues elicits all the classical cardinal signs of inflammation: redness, local heat, swelling and pain (115-117). In line with these symptoms, kinins are involved in cardiovascular regulation and mediation of inflammation and pain transmission.

Cardiovascular functions

Systemic injection or infusion of BK or LysBK elicits vasodilating and hypotensive effects in various animal species (33, 108, 118, 119) and man (120, 121), despite the fact that in vitro numerous major blood vessels respond to administration of kinins with constriction (6, 122). The vasorelaxant actions of kinins are mediated via endothelium-dependent and -independent mechanisms (34). However, the dominant net systemic effect is hypotension. The major endothelium-dependent vasorelaxation involves release of NO and secondary stimulation of cGMP production, although signaling via PGI₂ and endothelium-derived hyperpolarizing factors, e.g., epoxyeicosatrienoic acids, has also been postulated (34, 123). In contrast, DABK or LysDABK do not exhibit hypotensive actions without previous priming with inflammatory stimuli in most species, except dogs, due to the lack of constitutive expression of B₁ receptors, but display comparable hypotensive actions to BK after systemic inflammatory induction by LPS treatment (33, 108, 124, 125). Accordingly, B₂ receptor knockout mice and B₂ receptor antagonist-treated rats and mice have elevated basal blood pressure (126). Furthermore, they develop cardiomyopathy (127), are prone to further hypertension exacerbated by a high-salt diet (126, 128) and have disrupted preconditioning protection under conditions of repeated ischemia-reperfusion (129). These features clearly indicate the involvement of B₂ receptors in normal cardiovascular regulation and that long-term administration of B₂ receptor antagonists might have detrimental consequences with regard to cardiovascular risks. This is the main reason why B₂ receptor antagonists, although they could be antiinflammatory analgesics, have not gained importance in therapy as systemic treatment, except for the peptide antagonist icatibant (HOE-140) for the treatment of the acute condition hereditary- or ACE inhibitor-induced angioedema (130). Another B₂ receptor antagonist (anatibant, LF-16-0687) has also been developed for the treatment of an acute condition, traumatic brain injury (131). Furthermore, recently, the B₂ receptor antagonist MEN-16132 (fasitibant chloride hydrochloride) (132) has entered clinical development as a topical (intra-articular) treatment for osteoarthritis (OA) (ClinicalTrials.gov Identifier NCT01091116).

On the contrary, B₁ receptor knockout mice are healthy and normotensive (110). This is also in contrast with the phenotype of COX-2 knockout mice. It is worth noting that the COX-2 enzyme, like B₁ receptors, had been proposed to be constitutively expressed and thus not to participate in physiological but only in pathological processes. Nevertheless, COX-2 knockout mice exhibit elevated blood pressure, accelerated thrombogenesis (133), renal abnormalities, cardiac fibrosis and increased mortality (134), which is in line with the clinical observation of an increased incidence of myocardial infarction and stroke in patients treated with COX-2 inhibitors.

Kinins are believed to exert hypotensive and cardioprotective effects normally exclusively via the B₂ receptor. However, when B₂ receptors are pharmacologically or genetically ablated, the B₁ receptor under-

goes a compensatory overexpression and may take over their role (135, 136), suggesting that simultaneous blockade of both B₂ and B₁ receptors might cause more adverse cardiovascular effects than blocking B₂ receptors alone.

The major interest of researchers and drug companies turned in the last decade to the B₁ receptor and its antagonism as a promising drug target due to its special inducible nature (54, 83, 137, 138). However, in the context of therapeutic utility, it is not enough that B₁ receptors are not involved in physiological processes if B₁ receptor antagonists have detrimental consequences in pathological situations when B₁ receptors are induced. Therefore, numerous studies have been aimed at assessing the function of B₁ receptors in pathological situations by administering B₁ receptor antagonists or knock-out of B₁ receptors. The results are equivocal, with more hints for “salutary” than for “detrimental” effects, as summarized in Table III.

The controversies may be related to the fact that in most cases, when the inflammation itself is pathogenic and destructive, B₁ receptor-mediated reduction of inflammation is salutary, but in some severe conditions B₁ receptors may contribute to tissue reparation, which might be impaired by B₁ receptor ablation. It is worth noting that all the detrimental effects were seen in very severe conditions. The most troublesome finding in the context of side effect liability of B₁ receptor antagonists was enhanced atherosclerosis (154). However, this observation was also made in a seriously challenged condition, i.e., high-fat diet and knockout of apolipoprotein E (Apo-

E). Bradykinin B₁ receptors are upregulated in atherosclerotic plaques of human blood vessels and it has been proposed that inflammation, B₁ receptor stimulation and leukocyte migration may play a pathogenic role in early stages of atherosclerosis, suggesting that B₁ receptor antagonists may be useful for the prevention of atherosclerosis (157, 158). To sum up, B₁ receptor antagonism may be a two-edged sword with beneficial effects in early stages of atherosclerosis or when an inflammatory condition, such as rheumatoid arthritis (RA) (159), is the main pathogenic factor, but may be detrimental in severely progressed stages, or when serious dyslipidemia is the main pathogenic factor.

Central B₁ receptors also appear to be involved in pathological cardiovascular regulation. Bradykinin B₁ receptors are upregulated and kinin concentrations are increased in the hypothalamus of spontaneously hypertensive rats (SHR; a model of essential hypertension) (160, 161). Intracerebroventricular (i.c.v.) injection of LysDABK increases the blood pressure of both normal rats (Wistar-Kyoto, WKY) and SHR, which is in line with the constitutive expression of centrally located B₁ receptors (162). However, i.c.v. administration of B₁ receptor antagonists (DALBK or R-715) or antisense oligonucleotides for B₁ receptors, or systemic administration of brain-penetrating B₁ receptor antagonists (SSR-240612 or LF-22-0542), but not non-penetrating antagonists (R-715), markedly reduced blood pressure in hypertensive (either SHR or diabetes-induced), but not in normotensive control (WKY) rats (98, 100, 162, 163). These data sug-

Table III. Salutary and detrimental effects of pharmacological or genetic ablation of B₁ receptors in models of pathological conditions.

Ablation	Species	Condition/finding
Salutary effects		
KO	Mouse	Reduced infarct size in I/R injury of isolated heart (in vitro) (139)
KO	Mouse	Reduced myocardial infarct size following I/R injury (in vivo) (140)
KO & B1a	Mouse	Protection (reduced infarct) in a transient focal brain ischemia model (141)
KO & B1a	Mouse	Protection (including preserved renal function) against renal I/R injury (142, 143)
B1a	Mouse	Protection (including glomerular and tubular lesions, renal fibrosis, renal function) in nephrotoxic serum-induced glomerulonephritis model (144)
KO & B1a	Mouse	Protection against ureteral obstruction-induced nephropathy and fibrosis (145, 146)
B1a	Rat	Protection against I/R bowel injury and associated hypotension (147)
KO	Mouse	Attenuation of streptozotocin-induced diabetic cardiomyopathy (including improved systolic and diastolic function) (148)
KO	Mouse	Reduced (but B ₂ KO-enhanced) the hypotensive effects in lipopolysaccharide-induced model of endotoxemic shock (149)
B1a	Rat	Reduced plasma extravasation (microvascular permeability in numerous tissues including retina) in streptozotocin-induced diabetic vasculopathy (150-152)
Detrimental effects		
KO & B1a	Mouse	Revascularization and recovery of blood flow blunted after excision of femoral artery (153)
KO	Mouse	Enhanced formation of atherosclerotic plaques in animals provoked by cholesterol rich-diet and deletion of apolipoprotein E gene (154) (N.B. These animals show considerable damage without deleting the B ₁ receptor)
B1a	Rat	Reduced survival rate following pancreatitis induced by I/R (155)
KO	Mouse	Enhanced (but B ₂ KO-reduced) the hypotensive effect and mortality in lipopolysaccharide-induced model of endotoxemic shock (156)

KO, mice born with deleted bradykinin B₁ receptor gene (^{-/-}) compared to wild-type (^{+/+}); B1a, treatment with B₁ receptor antagonist compared to vehicle.

gest a beneficial (antihypertensive) rather than an adverse role for B₁ receptor antagonists with central disposition in pathological states of hypertension.

Overall, the above data, including studies with normal knockout animals, as well as those challenged in various disease models, suggest that the cardiovascular safety profile of B₁ receptor antagonists in a long-term treatment setting may be better than that of COX-2 inhibitors or B₂ receptor antagonists. In our view, there is a good chance that in a diseased population with some inflammatory condition a B₁ receptor antagonist might exhibit a cardiovascular risk-reducing effect. However, in conditions with serious damage, where healing via a proliferative process, e.g., fibrosis or revascularization, may be needed, avoiding treatment with B₁ receptor antagonists might be prudent. Ultimately, cardiovascular event outcome data from large clinical trials with B₁ receptor antagonists should give us definite answers.

Role in inflammation

Inflammation occurs as a response to infection or tissue injury and involves a highly complex interplay between multiple factors at the humoral, cellular and neuronal level. During inflammation, besides activation of the kallikrein-kinin system, other inflammatory mediators, e.g., cytokines, prostanoids, histamine, neuropeptides and NO, are released from various sources and comprise the “inflammatory soup”. The released mediators generate the symptoms of inflammation, i.e., vasodilation, extravasation and pain, as well as attract inflammatory cells to aid in elimination of microbes and tissue debris, and also to potentiate tissue repair. Kinins have been implicated in the release of all the above-mentioned mediators, but there are certainly redundant proinflammatory pathways. Due to the inducible nature of the B₁ receptor, the B₁ receptor agonist kinins are involved in positive feedback loops, such as cytokine > B₁ receptor induction and stimulation > chemotaxis and cellular response > cytokine and prostanoid release. Therefore, B₁ receptor antagonists can attenuate but do not completely block the inflammatory response. Both BK and DABK can increase the permeability of blood vessels and rapidly cause edema via stimulation of B₂ and B₁ receptors, respectively (164). Increased spontaneous and B₁ receptor agonist-induced extravasation is present, for example, in streptozotocin diabetic rats, and this can be almost completely reversed by chronic treatment with the B₁ receptor antagonists R-715 and R-954 (165, 166). Stimulation by B₁ receptors of fibroblasts leads to increased collagen synthesis (85) and B₁ receptor ablation prevents pathological fibrosis (145, 146, 148). Although fibrosis may be a reparative process after serious tissue injury, it is also an adverse consequence of chronic inflammation, for example, in the lung (167). In conclusion, mitigation of inflammation with B₁ receptor antagonists is a potential therapeutic opportunity for several indications, such as inflammatory bowel disease (168, 169). However, more mechanisms and opportunities related to potential antiinflammatory use of B₁ receptor antagonists are discussed elsewhere (54).

Role in pain transmission

Pain is an unpleasant sensory and emotional experience associated with actual or potential tissue damage (170). Pain sensation is normally transmitted and processed by pain pathways, which include various receptors (“nociceptors”) sensitive to mechanical, chemical

and thermal stimulus modalities, afferent nerve fibers with their cell bodies in the sensory ganglia (e.g., dorsal root or trigeminal ganglia), spinal cord neurons, and a complex network of spinal and supraspinal connections (“the pain matrix”) that processes and regulates the sensory and emotional experience and effector responses. Pain is a physiological process that serves the obvious function of avoiding damage. However, under pathological conditions, such as inflammation or nerve injury, the pain often outlives its usefulness as a warning system and instead becomes chronic and debilitating. This transition to a chronic phase involves changes within the spinal cord and brain, but there is also remarkable modulation where pain messages are initiated, i.e., at the level of the primary sensory neurons and their most distal structures, the nociceptors (171). Thus, pathological dysfunction of the system leads to peripheral and/or central sensitization, which manifests as spontaneous pain sensation, i.e., pain without stimuli, and exaggerated sensitivity to normally nonpainful innocuous stimuli, called “allodynia”, or to painful stimuli, called “hyperalgesia”. It is a serious gap between pain states in humans and pain research in animal models that a good measure of pain sensation does not exist and we usually rely on motor responses, i.e., nocifensive behaviors such as paw flinching or licking, withdrawal responses or vocalizations. In preclinical pain studies, depending on the method of stimulation, thermal hyperalgesia/allodynia (warm/hot stimuli), mechanical or tactile hyperalgesia/allodynia, cold allodynia or chemically induced nocifensive behaviors are investigated.

Bradykinin has long been known as an algogenic substance in humans when applied onto exposed tissues (“blister base”) or injected or microdialyzed into tissues (115, 117, 172, 173). This effect can be antagonized by a B₂ receptor antagonist (115). Experimental results indicate the same in animals, i.e., injection of B₂ receptor agonists (BK or [Tyr8]-BK) into the paw elicits nocifensive behaviors in naive mice (42) or rats (104). On the contrary, the B₁ receptor agonists LysDABK, microdialyzed into the skin in humans (173), or DABK, injected in rodents (42, 104, 174, 175), do not elicit significant pain or nocifensive behaviors in normal subjects. However, ample evidence suggests that after inflammatory stimuli and various neuropathic conditions, B₁ receptors are upregulated in the affected skin, afferent nerves, dorsal root ganglion (DRG) neurons, and a strong algogenic response appears upon application of B₁ receptor agonist peptide(s) in the form of nocifensive behaviors and hyperalgesia in animals (42, 104, 174, 175) and increased pain scoring in humans (173). The DABK-induced pain can be abolished by B₁ receptor antagonists applied either topically (175) or systemically (104). These functional pharmacodynamic results, in terms of pain perception, are consistent with the notion that B₂ receptors are constitutively present in healthy tissues, whereas B₁ receptors are inducible. On the other hand, when injected intrathecally, both BK and DABK elicited hyperalgesia (176, 177), a finding which might be consistent with constitutive expression of B₁ receptors in the spinal cord of naive animals (39). However, it should be noted that an intrathecally administered substance can also reach the dorsal root ganglia (178), where constitutive expression has also been demonstrated (37, 38, 41, 78, 177).

Genetic ablation of bradykinin B₁ receptors in mice causes slight hypoalgesia, as detected by increased latencies either in hot plate (110) or tail immersion flick (101) tests, but only in a limited stimulating temperature range (52–55.5 °C), i.e., mildly painful thermal stimulation. Testing thermal hyperalgesia by radiant heat or tactile

hyperalgesia by withdrawal threshold testing with von Frey filaments did not show any analgesic effect (101). This mild hypoalgesic effect in mice, to the best of our knowledge, has not been demonstrated with any B₁ receptor antagonist. However, in rats, in a hot plate test at 52 °C, one study demonstrated increased latency after administration of the reputedly CNS-penetrating B₁ receptor antagonist LF-22-0542 (101). Bradykinin B₁ receptor knockout mice exhibit attenuated nocifensive behavior compared to wild-type controls in the formalin test modeling tonic pain (101, 110). The pain induced by plantar injection of formalin has two characteristic phases, with an immediate appearance of nocifensive behavior (first phase: 0-5 minutes post-injection) followed by an apparent relief (around 10-15 minutes), and a second exacerbation of pain (15-40 minutes) (179). Apparently, both the first and second phases of formalin-induced pain were attenuated by approximately 50% in B₁ receptor-ablated mice (110). Studies with antagonists in rats and mice, including DALBK administered topically, systemically or intrathecally (102, 176, 180, 181), as well as SSR-240612 and LF-22-0542 administered systemically (99, 101), confirm a role for B₁ receptors in mediating formalin-induced pain. Interestingly, intrathecally administered DALBK attenuated only the second phase but not the first phase in mice (176). Genetic ablation or pharmacological blockade of B₁ receptors in mice by LF-22-0542 also attenuated the nocifensive behavior in the i.p. acetic acid-induced writhing test (101). The mild thermal hypoalgesia, the first-phase formalin-inhibitory effects and inhibition of writhing by B₁ receptor ablation suggest some role for B₁ receptors in acute nociception, which is independent from B₁ receptor induction. Whether this role is associated with the low level of constitutive expression of B₁ receptors in sensory neurons or in other cells releasing algogenic substances, or centrally located B₁ receptors play a role, remains elusive. However, these effects on acute nociception do not yield a strong analgesic action in acute pain.

The efficacy of B₁ receptor antagonists in inflammatory pain and hyperalgesia is well established. Several studies indicated mitigation of the role of B₂ receptors and involvement of that of B₁ receptors in conditions associated with inflammation (176, 182-185). B₁ receptor antagonists inhibit the hyperalgesia following acute inflammation induced by carrageenan (89, 101, 102, 186), which usually resolves within 24-48 hours, as well as the more chronic inflammation caused by CFA (109, 114, 176, 177, 184), which –depending on dose– lasts from 3 days to several weeks. The role of B₁ receptors in these inflammatory hyperalgesic states is supported by B₁ receptor knockout mouse data with measurement of either mechanical or thermal hyperalgesia (101, 114, 184). Bradykinin B₁ receptor antagonists are also effective pain relievers after several other inflammatory stimuli, such as injection of cytokines or zymosan, with similar efficacy to the NSAIDs diclofenac or indomethacin (103, 175). Accordingly, B₁ receptor antagonists hold promise as therapeutics for the treatment of chronic inflammatory conditions, particularly arthritis (54, 185, 187).

Nevertheless, until recently, very few studies investigated the efficacy of a B₁ receptor antagonist in animal models of genuine arthritis, i.e., monoarthritis elicited by CFA injection into the ankle or knee joint (102, 185, 188), not simply into the paw, or polyarthritis models, such as peptidoglycan-induced polyarthritis (189). Furthermore, only the effect of the partial agonist DALBK was tested in rats, which has its limitations, as discussed above. However, we recently published find-

ings showing that the novel nonpeptide peripherally acting B₁ receptor antagonist RGH-478 (0.3-3 mg/kg p.o. b.i.d.) effectively alleviated CFA-induced monoarthritic pain in both rats and rabbits, as assessed by the incapacitance test measuring the weight-bearing propensity of the affected leg (107). The antagonist was administered in an established subacute or chronic state of arthritis 3 and 11 days after CFA injection to rats and rabbits, respectively. The alleviation occurred immediately after the first dose and sustained pain relief developed in both species upon repeated dosing, which was equal or nonsignificantly superior compared to relevant doses of diclofenac and COX-2 inhibitors. The CFA model is considered to be a model of rheumatoid arthritis (RA). Although, osteoarthritis (OA) is also a plausible indication for B₁ receptor antagonists, we could not find any data in the public domain testing the effect of a B₁ receptor antagonist or genetic B₁ receptor ablation in an animal model of osteoarthritis. On the contrary, the efficacy of an intra-articularly administered B₂ receptor antagonist (MEN-16132) has been demonstrated in the monosodium iodoacetate-induced model of OA (132). The kallikrein-kinin system is activated and produces increased amounts of kinins in arthritis patients with various etiologies, including RA, OA, psoriatic arthritis and gout (30, 190, 191). Both B₁ and B₂ receptors are expressed in synoviocytes and endothelial cells in the synovial membrane of normal subjects and RA patients. Furthermore, in RA patients both B₁ and B₂ receptors are markedly upregulated in circulating neutrophil granulocytes, and even more extensively in neutrophils in the synovial fluid (71, 192). Therefore, the involvement of stimulation of both B₁ and B₂ receptors in arthritic inflammation and pain is likely not only in animal models, but also in human arthritis.

The peptide antagonist DALBK and the nonpeptide antagonist LF-22-0542 were shown to attenuate almost equally the nocifensive behaviors of mice in an experimental model of bone cancer pain (193). From the efficacy of the peptide it might be concluded that this analgesic effect is mediated by peripheral receptors. A caveat to this conclusion is that the dose of DALBK was rather high (10 µmol/kg s.c.), and thus its specificity as a selective antagonist of peripheral B₁ receptors may be challenged.

Painful neuropathies may result from nerve injury or damage of nerves elicited either by metabolic disorders, first of all diabetes, or by chronic treatment with neurotoxic drugs. The most widely used animal models of nerve injury include chronic constriction injury (CCI) (194) or partial nerve ligation (PNL) (195) of the sciatic nerve and the spinal nerve ligation (SNL) (196) model. Pharmacological or genetic ablation of B₁ receptors has analgesic effects on some aspects of injury-induced neuropathic pain. However, published results are controversial, probably due to differences in model conditions and pain modalities tested, as well as the involvement of peripheral neurogenic inflammatory components in the different models. Genetic ablation of B₁ receptors did not have any effect on the development of mechanical hyperalgesia in PNL (114) or tactile allodynia in the SNL model of neuropathic pain in mice (101). However, thermal hyperalgesia was alleviated by the genetic ablation of B₁ receptors in mice, and these findings, i.e., lack of effect on tactile allodynia but efficacy against thermal hyperalgesia, were further supported by pharmacological inhibition with LF-22-0542 in the SNL model in rats (101). In contrast, complete abolishment of both thermal hyperalgesia and tactile allodynia in B₁ receptor knockout mice was reported in the PNL model (174). However, in a rat PNL model, again, efficacy for LF-22-

0542 on thermal hyperalgesia, but not on mechanical and cold allodynia, was reported (41). Perhaps a species difference in the regulation of the expression of bradykinin receptors may account for this latter discrepancy. Namely, opposite changes –downregulation and upregulation– of B₂ receptors were observed in sensory neurons of mice (42) and rats (41), respectively, with simultaneous upregulation of B₁ receptors in both species following PNL surgery. To further the confusion, partial reversal of both cold allodynia and thermal hyperalgesia with relevant low doses of DALBK (with a bell-shaped dose–response relationship peaking at 100 nmol/kg i.p.) were observed in a rat SNL model. A small but statistically significant effect on mechanical allodynia was also observed in this study. In this rat model, upregulation of both B₁ and B₂ receptors was also observed in the skin and spinal nerves (104). There are at least three studies suggesting efficacy for B₁ receptor antagonists in the CCI model of neuropathic pain in rats. The nonpeptide B₁ receptor antagonist SSR-240612 caused dose-dependent reversal of thermal hyperalgesia in CCI rats. However, the doses needed for efficacy (20 and 30 mg/kg p.o.) in the CCI model were higher than those needed for maximum efficacy in inflammatory models or DABK-induced edema (3–10 mg/kg p.o.). Although the relevance of a study showing reversal of thermal hyperalgesia with extremely high single doses of DALBK (1–100 µmol/kg i.p.) may be questioned (197), another study with chronic (7-day) infusion of low doses (10 nmol/kg/day i.v.) of DALBK (198) strengthens the notion that B₁ receptor antagonists are also effective in alleviating neuropathic pain in the CCI model. In this latter study, both thermal hyperalgesia and tactile allodynia were completely reversed after 7 days of treatment. The results summarized above are quite consistent in that B₁ receptors, which are upregulated in the skin, nerves, sensory ganglia and spinal cord, are strongly involved in mediating thermal hyperalgesia following nerve injuries, but their involvement in other hyperalgesia modality domains is more controversial. The studies with genetic ablation of B₁ receptors do not differentiate between peripheral or central sites of involvement. The two nonpeptide B₁ receptor antagonists SSR-240612 and LF-22-0542 have been proposed to penetrate into the CNS. However, the peptide antagonist DALBK is believed not to enter the brain and spinal cord. Therefore, its anti-hyperalgesic efficacy could be an argument for the involvement of peripheral B₁ receptors in these states. On the other hand, since it is a partial agonist, its stimulating effect may also interfere with the hyperalgesia tests. Therefore, further studies with pure peripheral full B₁ receptor antagonists, such as R-715, in rodents would be needed to clarify the contribution of peripheral and central B₁ receptors to hyperalgesia in neuropathic pain conditions.

In a recent study, the involvement of different sites in a novel neuropathic pain model, the brachial plexus avulsion (ABP) test, was investigated and the effects of genetic ablation and pharmacological blockade of B₁ and B₂ receptors were tested (199). An interesting feature of this model is that the injured nerve innervates the forelimbs, but the thermal hyperalgesia and tactile allodynia (“hyperexcitability”) are measured from the ipsilateral hindlimbs, indicating involvement of at least long intersegmental and/or supraspinal integration. The tactile allodynia develops rapidly within 4 days and remains sustained for more than 80 days after surgery, whereas the thermal hyperalgesia lasts for about 20 days. Studies with knockout mice showed complete prevention of the allodynia and hyperalgesia

by B₁, but not by B₂ receptor ablation compared to wild-type animals. Studies with the B₁ receptor antagonists SSR-240612 and R-715 administered by local (surgery site), systemic (low doses: 0.3 and 0.5 mg/kg i.p., respectively), intrathecal and i.c.v. routes showed that, at the early stages (0–10 days post-surgery), local, systemic or lumbar intrathecal administration of either antagonist effectively alleviated hyperalgesia/allodynia, suggesting involvement of B₁ receptors at peripheral, DRG and/or spinal cord sites, whereas the smaller effect after i.c.v. administration suggested less involvement of supraspinal (brain) B₁ receptors. However, at a later stage, 30 days post-surgery, only i.c.v. administered antagonists proved effective, suggesting that at this stage only central supraspinal B₁ receptors are involved in the maintenance of hyperexcitability. These data also suggest that SSR-240612 at the low systemic dose applied, which is effective in inflammatory pain (99), does not provide sufficient central B₁ receptor blockade to alleviate this chronic pain state.

The B₁ receptor antagonist DALBK, although at a high dose of 1 µmol/kg i.p. in rats, was found to attenuate orofacial thermal hyperalgesia in a trigeminal pain model (200). To sum up, a large body of evidence suggests analgesic efficacy for B₁ receptor antagonists against some aspects of nerve injury-induced neuropathic pain. The results are more unequivocal for thermal hyperalgesia, but controversial for mechanical and cold allodynia/hyperalgesia. Probably some effects are mediated by peripheral B₁ receptors, but centrally acting B₁ receptor antagonists might have a broader analgesic profile affecting some aspects of central sensitization.

The potential efficacy of B₁ receptor antagonists in painful diabetic neuropathy is a special issue. Diabetes is associated with vascular permeability changes leading to many complications, including nephropathy, retinopathy, hypertension, hyperalgesia and neuropathy. A multitude of data has demonstrated the induction of B₁ receptors in various tissues and cell types, including macrophages, fibroblasts, blood vessels, kidneys, spinal cord and brain, either by morphological or functional pharmacological approaches in rodent models of diabetes (164, 201–207). Various B₁ receptor antagonists, including the probably only peripherally acting R-715, were found to alleviate thermal hyperalgesia and tactile and cold allodynia in rats and mice (98, 204, 208, 209). Most studies relate to the streptozotocin (STZ)-induced model of type 1 diabetes, which involves inflammation and causes hyperalgesia, and which is partly independent of hyperglycemia (210). However, the development of hyperalgesia, the involvement of B₁ receptors and the effectiveness of B₁ receptor antagonists were shown to be present also in non-STZ models of diabetes, such as the rat glucose feeding model of type 2 diabetes with insulin resistance (208), or genetic models, such as Zucker diabetic fatty rats (type 2) (210) and nonobese diabetic mice (type 1) (211). Bradykinin B₁ receptor antagonists either alleviated the developed hyperalgesia by acute administration (208, 210, 211), or attenuated/prevented the development of hyperalgesia by chronic administration (212, 213). These findings are also supported by B₁ receptor knockout mouse data (214). Hence, a great body of evidence suggests the potential utility of B₁ receptor antagonist drugs in the treatment of diabetic neuropathic pain, and in this case, peripherally acting B₁ receptor antagonists will probably be sufficiently effective. What is more, other complications, such as oxidative stress (213), vasculopathy and increased plasma extravasation

(150, 151), cardiomyopathy (148) and retinopathy (152, 166), are also thought to be alleviated by treatment with B₁ receptor antagonists, according to preclinical data. Therefore, painful diabetic neuropathy and other dreadful complications of this devastating disease are among the most promising indications for B₁ receptor antagonists.

COMPETITION AND PROGRESS IN B₁ RECEPTOR ANTAGONIST DRUG RESEARCH

The promise of potential utility in chronic pain and in a large number of other indications with an unmet medical need has compelled numerous pharmaceutical companies to engage in the difficult task of producing orally active nonpeptide B₁ receptor antagonists. Details of the medicinal chemistry of these efforts have been summarized in recent reviews (138, 215, 216). However, here we briefly summarize the competition in B₁ receptor antagonist research, presenting a representative set of compounds from published scientific and patent literature (Figs. 1 and 2). For most patent references, the reader is advised to access the above three medicinal chemistry reviews. Researchers at Merck, Novartis and Fournier have played a pioneering role in establishing the toolbox of bradykinin receptor research. However, the first patent on nonpeptide B₁ receptor antagonists was filed by sanofi in 1996 (217). Further optimization of that series led to the discovery of the dipeptidomimetic SSR-240612 (99, 218). Although according to a medicinal chemist's perspective it does not appear to be a "drug-like" compound, it proved to be orally active in pharmacodynamic tests (99). It is characterized by high molecular weight (757 Da), high lipophilicity and probably possesses significant metabolic liabilities associated with the naphthalene moiety. However, pharmacokinetic data for SSR-240612 have not been published. The compound reached phase II clinical development by the end of 2005 according to portfolio information released by sanofi-aventis, but development was halted for undisclosed reasons and the compound was removed from portfolio reports by 2008. Thus, SSR-240612 became a good "pharmacological tool". Furthermore, sanofi-aventis in conjunction with the acquired Fovea Pharmaceuticals is developing FOV-2304 (structure undisclosed) (152), a potent antagonist of bradykinin B₁ receptors, active as eye drops for the treatment of diabetic macular edema (Press Release, October 1, 2009). Sanofi's entrance with an aryl-sulfonamide scaffold was followed by a wave of 46 patent applications between 1999 and 2005 (priority dates) claiming nonpeptide B₁ receptor antagonists. Competitor companies in this race included Novartis, Pharmacopeia, Bayer, Merck (MSD), Fournier, Elan, Amgen and Boehringer. However, of these efforts, apparently only two compounds (MK-0686 and AMG-379) entered clinical development. A second wave of patenting activity emerged from 2006, with 39 disclosed patents filed by Boehringer, Neurogen, Grünenthal, Evotec, Jerini and Gedeon Richter between 2006 and 2009. Of these, a compound from Gedeon Richter is in clinical development, RGH-478 (structure undisclosed), which was optimized for oral efficacy in arthritis (107). It is very likely that more clinical entries will be revealed in the near future from this second wave of efforts.

Most of the patents were covering compounds with a similar aryl-sulfonamide-containing pharmacophore (Fig. 1). However, Pharmacopeia, Elan, Merck and Neurogen have also invented non-sulfon compounds that resemble the sulfonamide family (e.g., top

four structures in Fig. 2). Within this family, the molecular weights of the compounds that retained specificity remained high, mostly above 500 Da, which makes it difficult, although not impossible, to optimize a viable clinical candidate with good drug disposition and metabolic properties. Merck was the most dominant player in the field of B₁ receptor antagonists with 21 patent applications. In addition, Merck scientists invented a completely new type of structure, the biaryls, with potent B₁ receptor antagonist effects, a class that was recently followed by Jerini (bottom six structures in Figure 2).

As can be concluded from both patent literature and publications, Merck's priority driving the optimization was to find a brain-penetrating clinical candidate. For this reason, they were fine-tuning potency, selectivity, molecular size, physicochemical and pharmacokinetic properties, and, in addition to oral bioavailability, they particularly focused on eliminating efflux enzyme (P-glycoprotein) liability to enable brain penetration. The first fruit of these efforts was MK-0686 (see Fig. 2; molecular weight 487 Da), which reached phase II clinical trials. According to information on www.clinicaltrials.gov, a postoperative dental pain study (NCT00533403) was started and completed in June 2004 with MK-0686. Then, an osteoarthritis study (NCT00296569) was started in September 2005 and completed in May 2006. The third study was for post-herpetic neuralgia (NCT00282763) and started in December 2005, but was prematurely terminated by August 2006. From this timing, it might be speculated that some adverse features may have emerged, since lack of efficacy in one indication probably would not have affected expectations in another. The development of MK-0686 was halted but the reason was not disclosed. Nevertheless, MK-0686 did not seem to be an ideal drug candidate. Its *in vivo* characterization in preclinical tests was limited by the fact that it has potent activity at human B₁ receptors ($K_i = 0.4$ nM), but very weak activity at rat B₁ receptors ($K_i = 1646$ nM). However, the oral bioavailability of MK-0686 in rats was acceptable (34%). The only published pharmacodynamic characterization for analgesic efficacy was a study of CFA-induced allodynia in human B₁ receptor knock-in mice. In this study, MK-0686 (cited as Compound 13b) had an ED₅₀ value of 9.8 mg/kg *p.o.*, with maximum efficacy at around 60 mg/kg (109), which represents a surprisingly low *in vivo* potency for a compound with sub-nanomolar affinity. Although brain penetration in mice was much lower due to mouse P-glycoprotein liability than in monkeys (109), and presumably in man, this probably does not explain the low *in vivo* potency in an inflammatory pain test, where the efficacy may be mediated via peripheral B₁ receptors. Another issue with MK-0686 was its affinity for pregnane X receptors (NR1I2, PXR) and associated cytochrome P450 metabolic enzyme-inducing property (219, 220). The goal of further improvements in brain penetration and elimination of PXR liability gave rise to further optimization efforts (109, 219, 221-224) and claims about selection of further clinical candidate compounds (222, 224). However, the developmental fate of these Merck compounds remains elusive.

Disclosed information is scarce about the fate of efforts of other companies. However, these reflect the struggling with optimization of such peptidomimetic compounds also afflicted by species specificity. Amgen had a development candidate, AMG-379 (structure not disclosed), which was in phase I clinical trials in 2007 according to an internet-published agenda of a New Zealand ethics committee, but only as parenteral (intravenous and subcutaneous) administra-

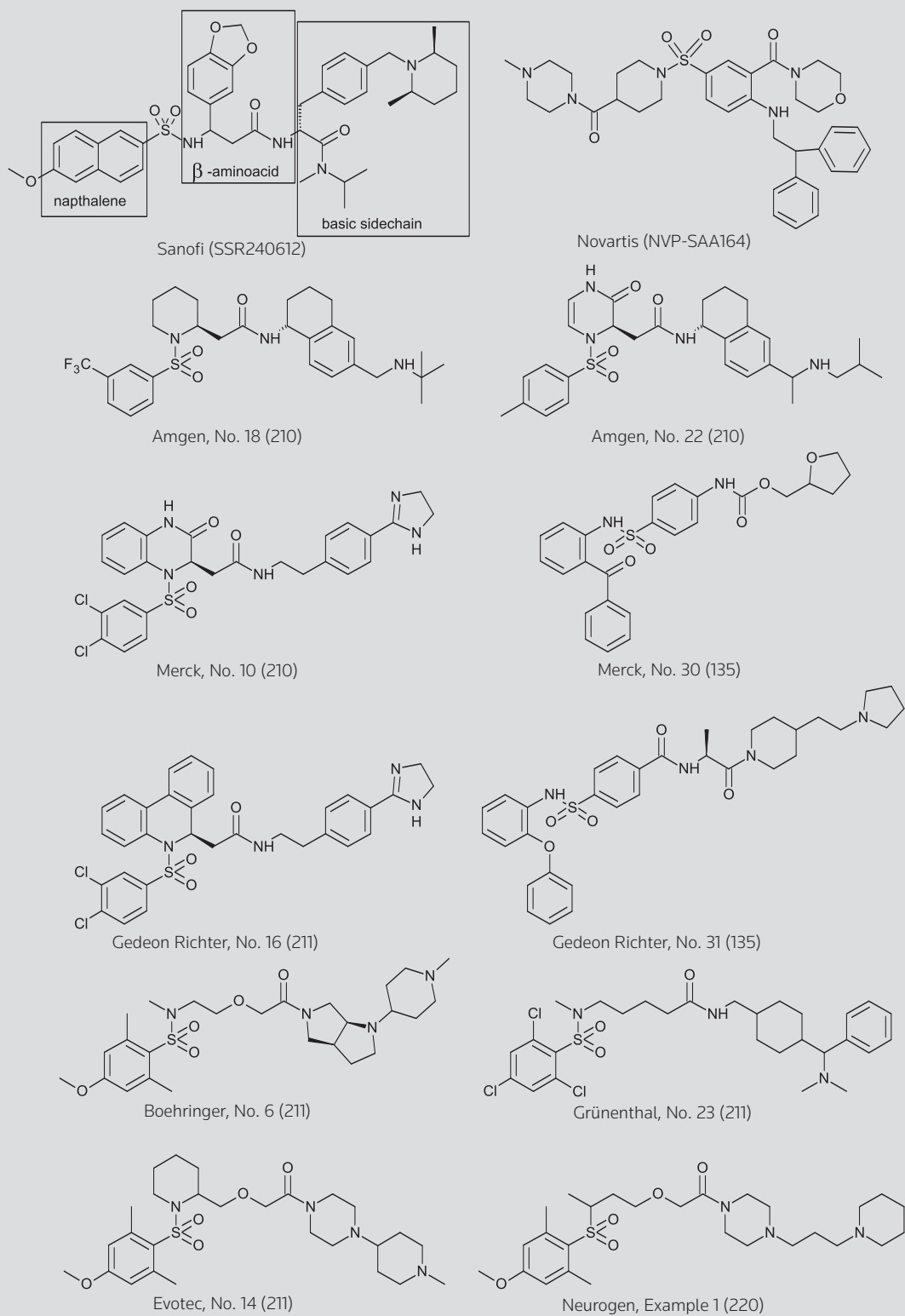
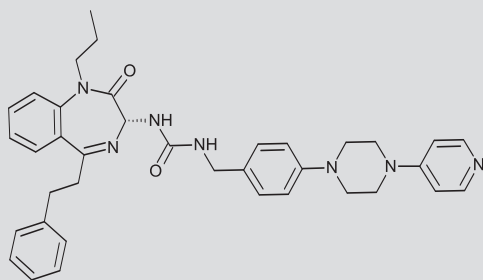
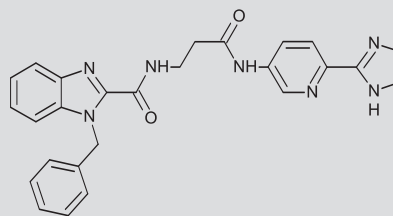


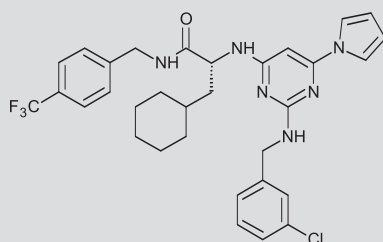
Figure 1. Representative B₁ antagonist arylsulfonamide structures from different companies.



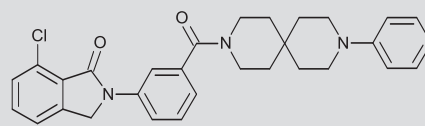
Merck, No. 33 (135)



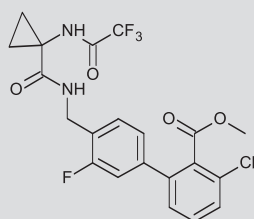
Neurogen, No. 34 (135)



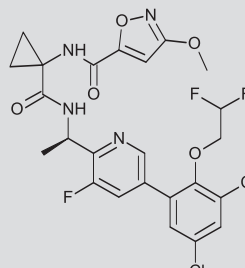
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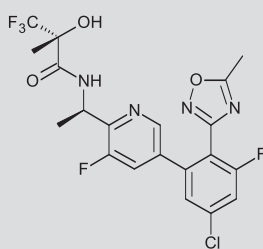
Elan, ELN441958



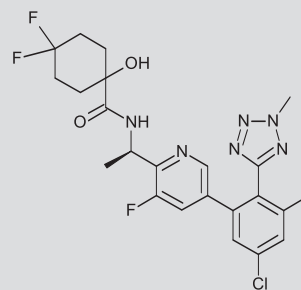
Merck, MK-0686



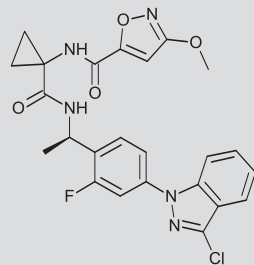
Merck, No. 7g (217)



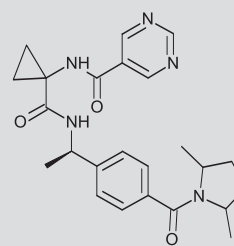
Merck, No. 21 (219)



Merck, No. 11i (218)



Merck, No. 10b (216)



Jerini, 2d (221)

Figure 2. Representative B₁ antagonist non-sulfonamide structures from different companies.

tion, suggesting that oral bioavailability was an issue. However, AMG-379 was dropped from Amgen's portfolio in 2008. Fournier apparently ended up with the research tool compound LF-22-0542 (see Table II), also with lack of sufficient oral bioavailability (101). Novartis also quit the field without a compound going to clinic but publishing favorable scientific results using B₁ receptor knockout and human B₁ receptor knock-in mice using a compound, NVP-SAA164, that was potent at human ($K_i = 8$ nM) but ineffective at rat and rabbit B₁ receptors (114). Elan researchers have also discovered and described a very interesting compound, but not free from issues. ELN-441958 had good oral bioavailability in both rats and monkeys and was highly potent at primate B₁ receptors ($K_b = 0.12$ and 0.24 nM, respectively, for human and monkey receptors) and also considerably potent at rodent receptors ($K_b = 1.5$ and 14 nM, respectively, for rat and mouse receptors). Nevertheless, analgesic effects were not demonstrated in rodents. Instead, a carrageenan-induced thermal hyperalgesia test was established in rhesus monkeys, which indicated analgesic effects only at surprisingly high doses (3–10 mg/kg s.c.) and plasma concentrations (ca. $1\text{--}5$ μ M). Furthermore, at 10 mg/kg, side effects, such as facial flushing, paling and sedation, were observed and the compound probably caused histamine release, which may also have confounded the analgesic profiling (89).

These findings point to the great challenge of optimizing nonpeptide B₁ receptor antagonists and that fulfilling crude pharmacokinetic criteria and in vitro potencies and liability panels may be insufficient for good candidate selection.

CONCLUDING REMARKS

Preclinical data with some privileged B₁ receptor antagonists in concert with B₁ receptor knockout animals suggest that the B₁ receptor is an attractive therapeutic target for both inflammatory and neuropathic pain. However, this has to be confirmed in clinical studies with suitable clinical candidate compounds. The selection of good clinical candidates must also be supported by good in vivo pharmacodynamic results, which warrant proper drug disposition properties and high safety margins, even if this is hindered by species differences. The low success rate may be attributed to multiple factors, such as the species discrepancies in early animal studies, poor ADME properties of some compounds, or unexpected toxicity in clinical studies (138). The only two compounds known to have reached the clinical stage beyond phase I (SSR-240612 and MK-0686) were probably suboptimal. However, it is strongly hoped that some new clinical candidates will emerge from the second wave of medicinal chemistry efforts, which will be suitable for clinical proof-of-concept studies.

Another critical issue is: Do we need a brain-penetrating compound? On the one hand, it appears that for several indications, e.g., inflammatory pain or diabetic complications, there is no need for brain penetration. Adding brain penetration to the wish list with high priority may necessitate compromise in other aspects of this complicated multifactorial optimization. Furthermore, our knowledge on the function of central B₁ receptors is still very scarce. Therefore, it might be prudent to go to the clinic first with compounds optimized for efficacy in peripheral indications/disease models, such as arthritis or diabetic complications, to exploit at least this kind of promise of B₁ receptor antagonists. On the other hand, there is no doubt that appropriately distributing centrally acting B₁ receptor antagonists

might have a broader analgesic profile, particularly in neuropathic pain. Nevertheless, it is also apparent that B₁ receptor antagonists and knockout animals address only some aspects of neuropathic pain. Therefore, to aim for centrally acting B₁ receptor antagonists may be a secondary approach that may carry more risks but may also hold more promise.

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DISCLOSURES

The authors state no conflicts of interest.

REFERENCES

- Moreau, M.E., Garbacki, N., Molinaro, G., Brown, N.J., Marceau, F., Adam, A. *The kallikrein-kinin system: Current and future pharmacological targets*. J Pharmacol Sci 2005, 99(1): 6–38.
- Zhao, Y., Qiu, Q., Mahdi, F., Shariat-Madar, Z., Rojkaer, R., Schmaier, A.H. *Assembly and activation of HK-PK complex on endothelial cells results in bradykinin liberation and NO formation*. Am J Physiol Heart Circ Physiol 2001, 280(4): H1821–9.
- Chai, K.X., Chen, L.M., Chao, J., Chao, L. *Kallistatin: A novel human serine proteinase inhibitor. Molecular cloning, tissue distribution, and expression in Escherichia coli*. J Biol Chem 1993, 268(32): 24498–505.
- Chao, J., Schmaier, A., Chen, L.M., Yang, Z., Chao, L. *Kallistatin, a novel human tissue kallikrein inhibitor: Levels in body fluids, blood cells, and tissues in health and disease*. J Lab Clin Med 1996, 127(6): 612–20.
- Regoli, D., Barabe, J., Park, W.K. *Receptors for bradykinin in rabbit aortae*. Can J Physiol Pharmacol 1977, 55(4): 855–67.
- Regoli, D., Barabe, J. *Pharmacology of bradykinin and related kinins*. Pharmacol Rev 1980, 32(1): 1–46.
- Hess, J.F., Borkowski, J.A., Stonesifer, G.Y., MacNeil, T., Strader, C.D., Ransom, R.W. *Cloning and pharmacological characterization of bradykinin receptors*. Braz J Med Biol Res 1994, 27(8): 1725–31.
- Hess, J.F., Derrick, A.W., MacNeil, T., Borkowski, J.A. *The agonist selectivity of a mouse B1 bradykinin receptor differs from human and rabbit B1 receptors*. Immunopharmacology 1996, 33(1–3): 1–8.
- McEachern, A.E., Shelton, E.R., Bhakta, S. et al. *Expression cloning of a rat B2 bradykinin receptor*. Proc Natl Acad Sci U S A 1991, 88(17): 7724–8.
- Ni, A., Chai, K.X., Chao, L., Chao, J. *Molecular cloning and expression of rat bradykinin B1 receptor*. Biochim Biophys Acta 1998, 1442(2–3): 177–85.
- Bachvarov, D.R., Saint-Jacques, E., Larrivee, J.F., Levesque, L., Rioux, F., Drapeau, G., Marceau, F. *Cloning and pharmacological characterization of the rabbit bradykinin B2 receptor*. J Pharmacol Exp Ther 1995, 275(3): 1623–30.
- MacNeil, T., Bieri, K.K., Menke, J.G., Hess, J.F. *Cloning and pharmacological characterization of a rabbit bradykinin B1 receptor*. Biochim Biophys Acta 1995, 1264(2): 223–8.
- Hess, J.F., Hey, P.J., Chen, T.B., O'Brien, J., Omalley, S.S., Pettibone, D.J., Chang, R.S. *Molecular cloning and pharmacological characterization of the canine B1 and B2 bradykinin receptors*. Biol Chem 2001, 382(1): 123–9.
- Eggerickx, D., Raspe, E., Bertrand, D., Vassart, G., Parmentier, M. *Molecular cloning, functional expression and pharmacological characteri-*

- zation of a human bradykinin B2 receptor gene. *Biochem Biophys Res Commun* 1992, 187(3): 1306-13.
15. Hess, J.F., Borkowski, J.A., Young, G.S., Strader, C.D., Ransom, R.W. Cloning and pharmacological characterization of a human bradykinin (BK-2) receptor. *Biochem Biophys Res Commun* 1992, 184(1): 260-8.
 16. Menke, J.G., Borkowski, J.A., Bierilo, K.K. et al. Expression cloning of a human B1 bradykinin receptor. *J Biol Chem* 1994, 269(34): 21583-6.
 17. Farmer, S.G., Burch, R.M., Meeker, S.A., Wilkins, D.E. Evidence for a pulmonary B3 bradykinin receptor. *Mol Pharmacol* 1989, 36(1): 1-8.
 18. Regoli, D., Nsa, A.S., Rizzi, A., Gobeil, F.J. Bradykinin receptors and their antagonists. *Eur J Pharmacol* 1998, 348(1): 1-10.
 19. Marceau, F., Hess, J.F., Bachvarov, D.R. The B1 receptors for kinins. *Pharmacol Rev* 1998, 50(3): 357-86.
 20. Erdos, E.G. Some old and some new ideas on kinin metabolism. *J Cardiovasc Pharmacol* 1990, 15(Suppl. 6): S20-S24.
 21. Pesquero, J.B., Pesquero, J.L., Oliveira, S.M., Roscher, A.A., Metzger, R., Ganten, D., Bader, M. Molecular cloning & functional characterization of a mouse bradykinin B1 receptor gene. *Biochem Biophys Res Commun* 1996, 224(1): 281.
 22. Hess, J.F., Hey, P.J., Chen, T.B., Pettibone, D.J., Chang, R.S. Molecular and pharmacological diversity of the kinin B1 receptor. *Int Immunopharmacol* 2002, 2(13-14): 1747-54.
 23. Bastian, S., Loillier, B., Paquet, J.L., Pruneau, D. Stable expression of human kinin B1 receptor in 293 cells: Pharmacological and functional characterization. *Br J Pharmacol* 1997, 122(2): 393-9.
 24. Paquet, J.L., Luccarini, J.M., Fouchet, C. et al. Pharmacological characterization of the bradykinin B2 receptor: Inter-species variability and dissociation between binding and functional responses. *Br J Pharmacol* 1999, 126(5): 1083-90.
 25. Simpson, P.B., Woollacott, A.J., Hill, R.G., Seabrook, G.R. Functional characterization of bradykinin analogues on recombinant human bradykinin B(1) and B(2) receptors. *Eur J Pharmacol* 2000, 392(1-2): 1-9.
 26. Gobeil, F., Pheng, L.H., Badini, I. et al. Receptors for kinins in the human isolated umbilical vein. *Br J Pharmacol* 1996, 118(2): 289-94.
 27. Decarie, A., Adam, A., Couture, R. Effects of captopril and Icatibant on bradykinin (BK) and des [Arg9] BK in carrageenan-induced edema. *Peptides* 1996, 17(6): 1009-15.
 28. Hamza, M., Wang, X.M., Adam, A., Brahim, J.S., Rowan, J.S., Carmona, G.N., Dionne, R.A. Kinin B1 receptors contributes to acute pain following minor surgery in humans. *Mol Pain* 2010, 6: 12.
 29. Raymond, P., Drapeau, G., Raut, R., Audet, R., Marceau, F., Ong, H., Adam, A. Quantification of des-Arg9-bradykinin using a chemiluminescence enzyme immunoassay: Application to its kinetic profile during plasma activation. *J Immunol Methods* 1995, 180(2): 247-57.
 30. Melmon, K.L., Webster, M.E., Goldfinger, S.E., Seegmiller, J.E. The presence of a kinin in inflammatory synovial effusion from arthritides of varying etiologies. *Arthritis Rheum* 1967, 10(1): 13-20.
 31. Decarie, A., Raymond, P., Gervais, N., Couture, R., Adam, A. Serum inter-species differences in metabolic pathways of bradykinin and [des-Arg9]BK: influence of enalaprilat. *Am J Physiol* 1996, 271(4, Pt. 2): H1340-7.
 32. Blais, C. Jr., Drapeau, G., Raymond, P., Lamontagne, D., Gervais, N., Venneman, I., Adam, A. Contribution of angiotensin-converting enzyme to the cardiac metabolism of bradykinin: An interspecies study. *Am J Physiol* 1997, 273(5, Pt. 2): H2263-71.
 33. Nakhostine, N., Ribuot, C., Lamontagne, D., Nadeau, R., Couture, R. Mediation by B1 and B2 receptors of vasodepressor responses to intravenously administered kinins in anaesthetized dogs. *Br J Pharmacol* 1993, 110(1): 71-6.
 34. Regoli, D., Pheng, L.H., Allogho, S.N., Nguyen, L., Gobeil, F. Receptors for kinins: From classical pharmacology to molecular biology. *Immunopharmacology* 1996, 33(1-3): 116-22.
 35. Rhaleb, N.E., Dion, S., Barabe, J., Rouissi, N., Jukic, D., Drapeau, G., Regoli, D. Receptors for kinins in isolated arterial vessels of dogs. *Eur J Pharmacol* 1989, 162(3): 419-27.
 36. Mahabeer, R., Naidoo, S., Raidoo, D.M. Detection of tissue kallikrein and kinin B1 and B2 receptor mRNAs in human brain by in situ RT-PCR. *Metab Brain Dis* 2000, 15(4): 325-35.
 37. Wotherspoon, G., Winter, J. Bradykinin B1 receptor is constitutively expressed in the rat sensory nervous system. *Neurosci Lett* 2000, 294(3): 175-8.
 38. Ma, Q.P. The expression of bradykinin B(1) receptors on primary sensory neurones that give rise to small caliber sciatic nerve fibres in rats. *Neuroscience* 2001, 107(4): 665-73.
 39. Ma, Q.P., Heavens, R. Basal expression of bradykinin B(1) receptor in the spinal cord in humans and rats. *Neuroreport* 2001, 12(11): 2311-4.
 40. Shughrue, P.J., Ky, B., Austin, C.P. Localization of B1 bradykinin receptor mRNA in the primate brain and spinal cord: An in situ hybridization study. *J Comp Neurol* 2003, 465(3): 372-84.
 41. Petcu, M., Dias, J.P., Ongali, B., Thibault, G., Neugebauer, W., Couture, R. Role of kinin B1 and B2 receptors in a rat model of neuropathic pain. *Int Immunopharmacol* 2008, 8(2): 188-96.
 42. Rashid, M.H., Inoue, M., Matsumoto, M., Ueda, H. Switching of bradykinin-mediated nociception following partial sciatic nerve injury in mice. *J Pharmacol Exp Ther* 2004, 308(3): 1158-64.
 43. Bouthillier, J., deBlois, D., Marceau, F. Studies on the induction of pharmacological responses to des-Arg9-bradykinin in vitro and in vivo. *Br J Pharmacol* 1987, 92(2): 257-64.
 44. Vellani, V., Zachrisson, O., McNaughton, P.A. Functional bradykinin B1 receptors are expressed in nociceptive neurones and are upregulated by the neurotrophin GDNF. *J Physiol* 2004, 560(Pt. 2): 391-401.
 45. Prat, A., Weinrib, L., Becher, B., Poirier, J., Duquette, P., Couture, R., Antel, J.P. Bradykinin B1 receptor expression and function on T lymphocytes in active multiple sclerosis. *Neurology* 1999, 53(9): 2087-92.
 46. Phagoo, S.B., Poole, S., Leeb-Lundberg, L.M. Autoregulation of bradykinin receptors: Agonists in the presence of interleukin-1beta shift the repertoire of receptor subtypes from B2 to B1 in human lung fibroblasts. *Mol Pharmacol* 1999, 56(2): 325-33.
 47. Haddad, E.B., Fox, A.J., Rousell, J., Burgess, G., McIntyre, P., Barnes, P.J., Chung, K.F. Post-transcriptional regulation of bradykinin B1 and B2 receptor gene expression in human lung fibroblasts by tumor necrosis factor-alpha: Modulation by dexamethasone. *Mol Pharmacol* 2000, 57(6): 1123-31.
 48. Hayashi, R., Yamashita, N., Matsui, S. et al. Bradykinin stimulates IL-6 and IL-8 production by human lung fibroblasts through ERK- and p38 MAPK-dependent mechanisms. *Eur Respir J* 2000, 16(3): 452-8.
 49. Phagoo, S.B., Reddi, K., Anderson, K.D., Leeb-Lundberg, L.M., Warburton, D. Bradykinin B1 receptor up-regulation by interleukin-1beta and B1 agonist occurs through independent and synergistic intracellular signaling mechanisms in human lung fibroblasts. *J Pharmacol Exp Ther* 2001, 298(1): 77-85.
 50. Tonussi, C.R., Ferreira, S.H. Bradykinin-induced knee joint incapacitation involves bradykinin B2 receptor mediated hyperalgesia and bradykinin B1 receptor-mediated nociception. *Eur J Pharmacol* 1997, 326(1): 61-5.
 51. Guevara-Lora, I., Florkowska, M., Kozik, A. Bradykinin-related peptides up-regulate the expression of kinin B1 and B2 receptor genes in human promonocytic cell line U937. *Acta Biochim Pol* 2009, 56(3): 515-22.

52. Schanstra, J.P., Bataille, E., Marin Castano, M.E. et al. *The B₁-agonist [des-Arg¹⁰]-kallidin activates transcription factor NF-kappaB and induces homologous upregulation of the bradykinin B₁-receptor in cultured human lung fibroblasts*. J Clin Invest 1998, 101(10): 2080-91.
53. Passos, G.F., Fernandes, E.S., Campos, M.M. et al. *Kinin B₁ receptor up-regulation after lipopolysaccharide administration: Role of proinflammatory cytokines and neutrophil influx*. J Immunol 2004, 172(3): 1839-47.
54. Calixto, J.B., Medeiros, R., Fernandes, E.S., Ferreira, J., Cabrini, D.A., Campos, M.M. *Kinin B₁ receptors: Key G-protein-coupled receptors and their role in inflammatory and painful processes*. Br J Pharmacol 2004, 143(7): 803-18.
55. Prado, G.N., Taylor, L., Zhou, X., Ricupero, D., Mierke, D.F., Polgar, P. *Mechanisms regulating the expression, self-maintenance, and signaling-function of the bradykinin B₂ and B₁ receptors*. J Cell Physiol 2002, 193(3): 275-86.
56. Campos, M.M., Souza, G.E., Calixto, J.B. *Upregulation of B₁ receptor mediating des-Arg⁹-BK-induced rat paw oedema by systemic treatment with bacterial endotoxin*. Br J Pharmacol 1996, 117(5): 793-8.
57. Tiffany, C.W., Burch, R.M. *Bradykinin stimulates tumor necrosis factor and interleukin-1 release from macrophages*. FEBS Lett 1989, 247(2): 189-92.
58. Sakamoto, W., Kaga, M., Handa, H. et al. *Bradykinin and Met-T-kinin-Leu stimulated PGE₂ production by rat macrophage and fibroblast*. Braz J Med Biol Res 1994, 27(8): 1923-7.
59. Galizzi, J.P., Bodinier, M.C., Chapelain, B. et al. *Up-regulation of [3H]-des-Arg¹⁰-kallidin binding to the bradykinin B₁ receptor by interleukin-1 beta in isolated smooth muscle cells: Correlation with B₁ agonist-induced PGI₂ production*. Br J Pharmacol 1994, 113(2): 389-94.
60. Lerner, U.H., Sahlberg, K., Ljunggren, O. *Thrombin and bradykinin enhance prostaglandin production in human peripheral blood monocytes*. J Oral Pathol Med 1989, 18(4): 246-50.
61. Sato, E., Koyama, S., Nomura, H., Kubo, K., Sekiguchi, M. *Bradykinin stimulates alveolar macrophages to release neutrophil, monocyte, and eosinophil chemotactic activity*. J Immunol 1996, 157(7): 3122-9.
62. Kuhr, F., Lowry, J., Zhang, Y., Brovkovich, V., Skidgel, R.A. *Differential regulation of inducible and endothelial nitric oxide synthase by kinin B₁ and B₂ receptors*. Neuropeptides 2010, 44(2): 145-54.
63. Mathis, S.A., Criscimagna, N.L., Leeb-Lundberg, L.M. *B₁ and B₂ kinin receptors mediate distinct patterns of intracellular Ca²⁺ signaling in single cultured vascular smooth muscle cells*. Mol Pharmacol 1996, 50(1): 128-39.
64. Meini, S., Lecci, A., Maggi, C.A. *The longitudinal muscle of rat ileum as a sensitive monoreceptor assay for bradykinin B₁ receptors*. Br J Pharmacol 1996, 117(8): 1619-24.
65. Nsa, A.S., Gobeil, F., Pheng, L.H., Nguyen, L., Neugebauer, W., Regoli, D. *Antagonists for kinin B₁ and B₂ receptors in the mouse*. Can J Physiol Pharmacol 1997, 75(6): 558-62.
66. Koyama, S., Sato, E., Numanami, H., Kubo, K., Nagai, S., Izumi, T. *Bradykinin stimulates lung fibroblasts to release neutrophil and monocyte chemotactic activity*. Am J Respir Cell Mol Biol 2000, 22(1): 75-84.
67. Brechter, A.B., Persson, E., Lundgren, I., Lerner, U.H. *Kinin B₁ and B₂ receptor expression in osteoblasts and fibroblasts is enhanced by interleukin-1 and tumour necrosis factor-alpha. Effects dependent on activation of NF-kappaB and MAP kinases*. Bone 2008, 43(1): 72-83.
68. Shirasaki, H., Kanaizumi, E., Himi, T. *Immunohistochemical localization of the bradykinin B₁ and B₂ receptors in human nasal mucosa*. Mediators Inflamm 2009, 102406.
69. Stadnicki, A., Pastucha, E., Nowaczyk, G. et al. *Immunolocalization and expression of kinin B_{1R} and B_{2R} receptors in human inflammatory bowel disease*. Am J Physiol Gastrointest Liver Physiol 2005, 289(2): G361-6.
70. Wang, D.Z., Song, Q., Chen, L.M., Chao, L., Chao, J. *Expression and cellular localization of tissue kallikrein-kinin system in human adrenal gland*. Am J Physiol 1996, 271(3, Pt. 2): F709-16.
71. Cassim, B., Mody, G., Bhoola, K. *Kallikrein cascade and cytokines in inflamed joints*. Pharmacol Ther 2002, 94(1-2): 1-34.
72. Bertram, C., Misso, N.L., Fogel-Petrovic, M., Figueroa, C., Thompson, P.J., Bhoola, K.D. *Comparison of kinin B₁(1) and B₂(2) receptor expression in neutrophils of asthmatic and non-asthmatic subjects*. Int Immunopharmacol 2007, 7(14): 1862-8.
73. Bertram, C.M., Misso, N.L., Fogel-Petrovic, M., Figueroa, C.D., Foster, P.S., Thompson, P.J., Bhoola, K.D. *Expression of kinin receptors on eosinophils: Comparison of asthmatic patients and healthy subjects*. J Leukoc Biol 2009, 85(3): 544-52.
74. Bertram, C.M., Baltic, S., Misso, N.L., Bhoola, K.D., Foster, P.S., Thompson, P.J., Fogel-Petrovic, M. *Expression of kinin B₁ and B₂ receptors in immature, monocyte-derived dendritic cells and bradykinin-mediated increase in intracellular Ca²⁺ and cell migration*. J Leukoc Biol 2007, 81(6): 1445-54.
75. Dlamini, Z., Bhoola, K.D. *Upregulation of tissue kallikrein, kinin B₁ receptor, and kinin B₂ receptor in mast and giant cells infiltrating oesophageal squamous cell carcinoma*. J Clin Pathol 2005, 58(9): 915-22.
76. Raidoo, D.M., Bhoola, K.D. *Kinin receptors on human neurones*. J Neuroimmunol 1997, 77(1): 39-44.
77. Ongali, B., Campos, M.M., Bregola, G. et al. *Autoradiographic analysis of rat brain kinin B₁ and B₂ receptors: Normal distribution and alterations induced by epilepsy*. J Comp Neurol 2003, 461(4): 506-19.
78. Ma, Q.P., Hill, R., Sirinathsinghji, D. *Basal expression of bradykinin B₁ receptor in peripheral sensory ganglia in the rat*. Neuroreport 2000, 11(18): 4003-5.
79. Seabrook, G.R., Bowery, B.J., Heavens, R. et al. *Expression of B₁ and B₂ bradykinin receptor mRNA and their functional roles in sympathetic ganglia and sensory dorsal root ganglia neurones from wild-type and B₂ receptor knockout mice*. Neuropharmacology 1997, 36(7): 1009-17.
80. Davis, C.L., Naeem, S., Phagoo, S.B., Campbell, E.A., Urban, L., Burgess, G.M. *B₁ bradykinin receptors and sensory neurones*. Br J Pharmacol 1996, 118(6): 1469-76.
81. Petho, G., Derow, A., Reeh, P.W. *Bradykinin-induced nociceptor sensitization to heat is mediated by cyclooxygenase products in isolated rat skin*. Eur J Neurosci 2001, 14(2): 210-8.
82. Raidoo, D.M., Bhoola, K.D. *Pathophysiology of the kallikrein-kinin system in mammalian nervous tissue*. Pharmacol Ther 1998, 79(2): 105-27.
83. Leeb-Lundberg, L.M., Marceau, F., Muller-Esterl, W., Pettibone, D.J., Zuraw, B.L. *International union of pharmacology. XLV. Classification of the kinin receptor family: From molecular mechanisms to pathophysiological consequences*. Pharmacol Rev 2005, 57(1): 27-77.
84. Ricupero, D.A., Romero, J.R., Rishikof, D.C., Goldstein, R.H. *Des-Arg¹⁰-kallidin engagement of the B₁ receptor stimulates type I collagen synthesis via stabilization of connective tissue growth factor mRNA*. J Biol Chem 2000, 275(17): 12475-80.
85. Goldstein, R.H., Wall, M. *Activation of protein formation and cell division by bradykinin and des-Arg⁹-bradykinin*. J Biol Chem 1984, 259(14): 9263-8.
86. Luksha, L., Agewall, S., Kublickiene, K. *Endothelium-derived hyperpolarizing factor in vascular physiology and cardiovascular disease*. Atherosclerosis 2009, 202(2): 330-44.
87. Kuhr, F.K., Zhang, Y., Brovkovich, V., Skidgel, R.A. *Beta-arrestin 2 is required for B₁ receptor-dependent post-translational activation of inducible nitric oxide synthase*. FASEB J 2010, 24(7): 2475-83.

88. Regoli, D., Rizzi, A., Perron, S.I., Gobeil, F. Jr. *Classification of kinin receptors*. Biol Chem 2001, 382(1): 31-5.
89. Hawkinson, J.E., Szoke, B.G., Garofalo, A.W. et al. *Pharmacological, pharmacokinetic, and primate analgesic efficacy profile of the novel bradykinin B1 receptor antagonist ELN441958*. J Pharmacol Exp Ther 2007, 322(2): 619-30.
90. Cote, J., Savard, M., Bovenzi, V. et al. *Novel kinin B1 receptor agonists with improved pharmacological profiles*. Peptides 2009, 30(4): 788-95.
91. MacNeil, T., Feighner, S., Hreniuk, D.L., Hess, J.F., Van der Ploeg, L.H. *Partial agonists and full antagonists at the human and murine bradykinin B1 receptors*. Can J Physiol Pharmacol 1997, 75(6): 735-40.
92. Nsa Allogho, S., Gobeil, F., Pheng, L.H., Nguyen, L., Neugebauer, W., Regoli, D. *Kinin B1 and B2 receptors in the mouse*. Can J Physiol Pharmacol 1995, 73(12): 1759-64.
93. Wirth, K., Breipohl, G., Stechl, J., Knolle, J., Henke, S., Scholkens, B. *DesArg9-D-Arg[Hyp3,Thi5,D-Tic7,Oic8]bradykinin (desArg10-[Hoe140]) is a potent bradykinin B1 receptor antagonist*. Eur J Pharmacol 1991, 205(2): 217-8.
94. Szabo, A., Helyes, Z., Sandor, K. et al. *Role of transient receptor potential vanilloid 1 receptors in adjuvant-induced chronic arthritis: In vivo study using gene-deficient mice*. J Pharmacol Exp Ther 2005, 314(1): 111-9.
95. Gobeil, F., Neugebauer, W., Filteau, C. et al. *Structure-activity studies of B1 receptor-related peptides. Antagonists*. Hypertension 1996, 28(5): 833-9.
96. Nsa Allogho S., Gobeil, F., Pheng, L.H., Nguyen, L., Neugebauer, W., Regoli, D. *Antagonists for kinin B1 and B2 receptors in the mouse*. Can J Physiol Pharmacol 1997, 75(6): 558-62.
97. Neugebauer, W., Blais, P.A., Halle, S., Filteau, C., Regoli, D., Gobeil, F. Jr. *Kinin B1 receptor antagonists with multi-enzymatic resistance properties*. Can J Physiol Pharmacol 2002, 80(4): 287-92.
98. Lungu, C., Dias, J.P., Franca, C.E., Ongali, B., Regoli, D., Moldovan, F., Couture, R. *Involvement of kinin B1 receptor and oxidative stress in sensory abnormalities and arterial hypertension in an experimental rat model of insulin resistance*. Neuropeptides 2007, 41(6): 375-87.
99. Gougat, J., Ferrari, B., Sarran, L. et al. *SSR240612 [(2R)-2-[[[(3R)-3-(1,3-benzodioxol-5-yl)-3-[[[(6-methoxy-2-naphthyl)sulfonyl]amino]propanoyl]-amino]-3-(4-[[[2R,6S]-2,6-dimethylpiperidinyl]methyl]phenyl)-N-isopropyl-N-methylpropanamide hydrochloride], a new nonpeptide antagonist of the bradykinin B1 receptor: Biochemical and pharmacological characterization*. J Pharmacol Exp Ther 2004, 309(2): 661-9.
100. De Brito, G.H., Carayon, P., Ferrari, B., Couture, R. *Contribution of the central dopaminergic system in the anti-hypertensive effect of kinin B1 receptor antagonists in two rat models of hypertension*. Neuropeptides 2010, 44(2): 191-8.
101. Porreca, F., Vanderah, T.W., Guo, W. et al. *Antinociceptive pharmacology of N-[[4-(4,5-dihydro-1H-imidazol-2-yl)phenyl]methyl]-2-[2-[[[(4-methoxy-2,6-dimethylphenyl)sulfonyl]methylamino]ethoxy]-N-methylacetamide, fumarate (LF22-0542), a novel nonpeptidic bradykinin B1 receptor antagonist*. J Pharmacol Exp Ther 2006, 318(1): 195-205.
102. Rupniak, N.M., Boyce, S., Webb, J.K. et al. *Effects of the bradykinin B1 receptor antagonist des-Arg9[Leu8]bradykinin and genetic disruption of the B2 receptor on nociception in rats and mice*. Pain 1997, 71(1): 89-97.
103. Belichard, P., Landry, M., Faye, P., Bachvarov, D.R., Bouthillier, J., Pruneau, D., Marceau, F. *Inflammatory hyperalgesia induced by zymosan in the plantar tissue of the rat: effect of kinin receptor antagonists*. Immunopharmacology 2000, 46(2): 139-47.
104. Werner, M.F., Kassuya, C.A., Ferreira, J., Zampronio, A.R., Calixto, J.B., Rae, G.A. *Peripheral kinin B(1) and B(2) receptor-operated mechanisms are implicated in neuropathic nociception induced by spinal nerve ligation in rats*. Neuropharmacology 2007, 53(1): 48-57.
105. Davis, A.J., Perkins, M.N. *Des-Arg9-BK-induced mechanical hyperalgesia and analgesia in the rat: Involvement of IL-1, prostaglandins and peripheral opioids*. Br J Pharmacol 1997, 120: 74P.
106. Mason, G.S., Cumberbatch, M.J., Hill, R.G., Rupniak, N.M. *The bradykinin B1 receptor antagonist B9858 inhibits a nociceptive spinal reflex in rabbits*. Can J Physiol Pharmacol 2002, 80(4): 264-8.
107. Farkas, S., Hornok, K., Kis-Varga, Á. et al. *Analgesic effects of a novel highly selective bradykinin B1 receptor antagonist in animal models of monoarthritis*. 12th World Cong Pain (IASP) (Aug 17-22, Glasgow) 2008, Abst PT213.
108. deBlois, D., Horlick, R.A. *Endotoxin sensitization to kinin B(1) receptor agonist in a non-human primate model: Haemodynamic and pro-inflammatory effects*. Br J Pharmacol 2001, 132(1): 327-35.
109. Kuduk, S.D., Di Marco, C.N., Chang, R.K. et al. *Development of orally bioavailable and CNS penetrant biphenylaminocyclopropane carboxamide bradykinin B1 receptor antagonists*. J Med Chem 2007, 50(2): 272-82.
110. Pesquero, J.B., Araujo, R.C., Heppenstall, P.A. et al. *Hypoalgesia and altered inflammatory responses in mice lacking kinin B1 receptors*. Proc Natl Acad Sci U S A 2000, 97(14): 8140-5.
111. Hess, J.F., Ransom, R.W., Zeng, Z. et al. *Generation and characterization of a human bradykinin receptor B1 transgenic rat as a pharmacodynamic model*. J Pharmacol Exp Ther 2004, 310(2): 488-97.
112. Su, D.S., Markowitz, M.K., Murphy, K.L. et al. *Development of an efficient and selective radioligand for bradykinin B1 receptor occupancy studies*. Bioorg Med Chem Lett 2004, 14(24): 6045-8.
113. Hess, J.F., Chen, R.Z., Hey, P. et al. *Generation and characterization of a humanized bradykinin B1 receptor mouse*. Biol Chem 2006, 387(2): 195-201.
114. Fox, A., Kaur, S., Li, B. et al. *Antihyperalgesic activity of a novel nonpeptide bradykinin B1 receptor antagonist in transgenic mice expressing the human B1 receptor*. Br J Pharmacol 2005, 144(7): 889-99.
115. Whalley, E.T., Clegg, S., Stewart, J.M., Vavrek, R.J. *The effect of kinin agonists and antagonists on the pain response of the human blister base*. Naunyn Schmiedebergs Arch Pharmacol 1987, 336(6): 652-5.
116. Marceau, F., Lussier, A., Regoli, D., Giroud, J.P. *Pharmacology of kinins: Their relevance to tissue injury and inflammation*. Gen Pharmacol 1983, 14(2): 209-29.
117. Armstrong, D., Jepson, J.B., Keele, C.A., Stewart, J.W. *Pain-producing substance in human inflammatory exudates and plasma*. J Physiol 1957, 135(2): 350-70.
118. Barabe, J., Marceau, F., Theriault, B., Drouin, J.N., Regoli, D. *Cardiovascular actions of kinins in the rabbit*. Can J Physiol Pharmacol 1979, 57(1): 78-91.
119. Griesbacher, T., Lembeck, F., Saria, A. *Effects of the bradykinin antagonist B4310 on smooth muscles and blood pressure in the rat, and its enzymatic degradation*. Br J Pharmacol 1989, 96(3): 531-8.
120. Norrby, C., Dencker, H., Lunderquist, A., Olin, T. *Superior mesenteric blood flow in man following injection of bradykinin and vasopressin into the superior mesenteric artery*. Acta Chir Scand 1975, 141(2): 119-28.
121. Saameli, K., Eskes, T.K. *Bradykinin and cardiovascular system: Estimation of half-life*. Am J Physiol 1962, 203: 261-5.
122. McLean, P.G., Perretti, M., Ahluwalia, A. *Kinin B(1) receptors and the cardiovascular system: Regulation of expression and function*. Cardiovasc Res 2000, 48(2): 194-210.
123. Campbell, W.B., Fleming, I. *Epoxyeicosatrienoic acids and endothelium-dependent responses*. Pflügers Arch 2010, 459(6): 881-95.
124. Regoli, D.C., Marceau, F., Lavigne, J. *Induction of beta 1-receptors for kinins in the rabbit by a bacterial lipopolysaccharide*. Eur J Pharmacol 1981, 71(1): 105-15.

125. Tokumasu, T., Ueno, A., Oh-ishi, S. A hypotensive response induced by des-Arg⁹-bradykinin in young brown/Norway rats pretreated with endotoxin. *Eur J Pharmacol* 1995, 274(1-3): 225-8.
126. Madeddu, P., Varoni, M.V., Palomba, D. et al. Cardiovascular phenotype of a mouse strain with disruption of bradykinin B₂-receptor gene. *Circulation* 1997, 96(10): 3570-8.
127. Emanuelli, C., Maestri, R., Corradi, D. et al. Dilated and failing cardiomyopathy in bradykinin B₂ receptor knockout mice. *Circulation* 1999, 100(23): 2359-65.
128. Alfie, M.E., Sigmon, D.H., Pomposiello, S.I., Carretero, O.A. Effect of high salt intake in mutant mice lacking bradykinin-B₂ receptors. *Hypertension* 1997, 29(1, Pt. 2): 483-7.
129. Yang, X.P., Liu, Y.H., Scicli, G.M., Webb, C.R., Carretero, O.A. Role of kinins in the cardioprotective effect of preconditioning: Study of myocardial ischemia/reperfusion injury in B₂ kinin receptor knockout mice and kininogen-deficient rats. *Hypertension* 1997, 30(3, Pt. 2): 735-40.
130. Sorbera, L.A., Fernandez-Forner, D., Bayes, M. Icatibant acetate: Treatment of hereditary angioedema, agent for liver cirrhosis, antiasthmatic drug, treatment of osteoarthritis pain, bradykinin B₂ antagonist. *Drugs Fut* 2006, 31(2): 101-6.
131. Zweckberger, K., Plesnila, N. Anatibant, a selective non-peptide bradykinin B₂ receptor antagonist, reduces intracranial hypertension and histopathological damage after experimental traumatic brain injury. *Neurosci Lett* 2009, 454(2): 115-7.
132. Cialdai, C., Giuliani, S., Valenti, C., Tramontana, M., Maggi, C.A. Effect of Intra-articular 4-(S)-amino-5-(4-[2,4-dichloro-3-(2,4-dimethyl-8-quinoloxymethyl)phenylsulfonamido]-tetrahydro-2H-4-pyran-2-yl)-piperazine-5-oxopentyl(trimethyl)ammonium chloride hydrochloride (MEN16132), a kinin B₂ receptor antagonist, on nociceptive response in monosodium iodoacetate-induced experimental osteoarthritis in rats. *J Pharmacol Exp Ther* 2009, 331(3): 1025-32.
133. Cheng, Y., Wang, M., Yu, Y., Lawson, J., Funk, C.D., FitzGerald, G.A. Cyclooxygenases, microsomal prostaglandin E synthase-1, and cardiovascular function. *J Clin Invest* 2006, 116(5): 1391-9.
134. Dinchuk, J.E., Car, B.D., Focht, R.J. et al. Renal abnormalities and an altered inflammatory response in mice lacking cyclooxygenase II. *Nature* 1995, 378(6555): 406-9.
135. Duka, A., Duka, I., Gao, G., Shenouda, S., Gavras, I., Gavras, H. Role of bradykinin B₁ and B₂ receptors in normal blood pressure regulation. *Am J Physiol Endocrinol Metab* 2006, 291(2): E268-74.
136. Duka, I., Duka, A., Kintsurashvili, E., Johns, C., Gavras, I., Gavras, H. Mechanisms mediating the vasoactive effects of the B₁ receptors of bradykinin. *Hypertension* 2003, 42(5): 1021-5.
137. Calixto, J.B., Cabrini, D.A., Ferreira, J., Campos, M.M. Kinins in pain and inflammation. *Pain* 2000, 87(1): 1-5.
138. Huang, H., Player, M.R. Bradykinin B₁ receptor antagonists as potential therapeutic agents for pain. *J Med Chem* 2010, 53(15): 5383-99.
139. Lagneux, C., Bader, M., Pesquero, J.B., Demenge, P., Ribuo, C. Detrimental implication of B₁ receptors in myocardial ischemia: Evidence from pharmacological blockade and gene knockout mice. *Int Immunopharmacol* 2002, 2(6): 815-22.
140. Yin, H., Chao, J., Bader, M., Chao, L. Differential role of kinin B₁ and B₂ receptors in ischemia-induced apoptosis and ventricular remodeling. *Peptides* 2007, 28(7): 1383-9.
141. Austinat, M., Braeuninger, S., Pesquero, J.B. et al. Blockade of bradykinin receptor B₁ but not bradykinin receptor B₂ provides protection from cerebral infarction and brain edema. *Stroke* 2009, 40(1): 285-93.
142. Wang, P.H., Cenedeze, M.A., Pesquero, J.B., Pacheco-Silva, A., Camara, N.O. Influence of bradykinin B₁ and B₂ receptors in the immune response triggered by renal ischemia-reperfusion injury. *Int Immunopharmacol* 2006, 6(13-14): 1960-5.
143. Wang, P.H., Campanholle, G., Cenedeze, M.A. et al. Bradykinin [corrected] B₁ receptor antagonism is beneficial in renal ischemia-reperfusion injury. *PLoS ONE* 2008, 3(8): e3050.
144. Klein, J., Gonzalez, J., Decramer, S. et al. Blockade of the kinin B₁ receptor ameliorates glomerulonephritis. *J Am Soc Nephrol* 2010, 21(7): 1157-64.
145. Klein, J., Gonzalez, J., Duchene, J. et al. Delayed blockade of the kinin B₁ receptor reduces renal inflammation and fibrosis in obstructive nephropathy. *FASEB J* 2009, 23(1): 134-42.
146. Wang, P.H., Cenedeze, M.A., Campanholle, G. et al. Deletion of bradykinin B₁ receptor reduces renal fibrosis. *Int Immunopharmacol* 2009, 9(6): 653-7.
147. Madeddu, P., Emanuelli, C., Bonaria, S.M. et al. Role of calcitonin gene-related peptide and kinins in post-ischemic intestinal reperfusion. *Peptides* 2001, 22(6): 915-22.
148. Westermann, D., Walther, T., Savvatis, K. et al. Gene deletion of the kinin receptor B₁ attenuates cardiac inflammation and fibrosis during the development of experimental diabetic cardiomyopathy. *Diabetes* 2009, 58(6): 1373-81.
149. Cayla, C., Todiras, M., Iliescu, R. et al. Mice deficient for both kinin receptors are normotensive and protected from endotoxin-induced hypotension. *FASEB J* 2007, 21(8): 1689-98.
150. Lawson, S.R., Gabra, B.H., Guerin, B., Neugebauer, W., Nantel, F., Battistini, B., Sirois, P. Enhanced dermal and retinal vascular permeability in streptozotocin-induced type 1 diabetes in Wistar rats: Blockade with a selective bradykinin B₁ receptor antagonist. *Regul Pept* 2005, 124(1-3): 221-4.
151. Lawson, S.R., Gabra, B.H., Nantel, F., Battistini, B., Sirois, P. Effects of a selective bradykinin B₁ receptor antagonist on increased plasma extravasation in streptozotocin-induced diabetic rats: Distinct vasculopathic profile of major key organs. *Eur J Pharmacol* 2005, 514(1): 69-78.
152. Pruneau, D., Belichard, P., Sahel, J.A., Combal, J.P. Targeting the kallikrein-kinin system as a new therapeutic approach to diabetic retinopathy. *Curr Opin Investig Drugs* 2010, 11(5): 507-14.
153. Emanuelli, C., Bonaria, S.M., Stacca, T. et al. Targeting kinin B₁ receptor for therapeutic neovascularization. *Circulation* 2002, 105(3): 360-6.
154. Merino, V.F., Todiras, M., Mori, M.A. et al. Predisposition to atherosclerosis and aortic aneurysms in mice deficient in kinin B₁ receptor and apolipoprotein E. *J Mol Med* 2009, 87(10): 953-63.
155. Kuebler, J.F., Schremmer-Danninger, E., Bhoola, K.D., Roscher, A.A., Messmer, K., Hoffmann, T.F. Kinin-B₁ receptors in ischaemia-induced pancreatitis: Functional importance and cellular localisation. *Biol Chem* 2003, 384(9): 1311-9.
156. Seguin, T., Buleon, M., Destrupe, M., Ranera, M.T., Couture, R., Girolami, J.P., Tack, I. Hemodynamic and renal involvement of B₁ and B₂ kinin receptors during the acute phase of endotoxin shock in mice. *Int Immunopharmacol* 2008, 8(2): 217-21.
157. Raidoo, D.M., Ramsaroop, R., Naidoo, S., Muller-Esterl, W., Bhoola, K.D. Kinin receptors in human vascular tissue: Their role in atheromatous disease. *Immunopharmacology* 1997, 36(2-3): 153-60.
158. Duchene, J., Ahluwalia, A. The kinin B₁ receptor and inflammation: New therapeutic target for cardiovascular disease. *Curr Opin Pharmacol* 2009, 9(2): 125-31.
159. Sodergren, A., Karp, K., Boman, K. et al. Atherosclerosis in early rheumatoid arthritis: very early endothelial activation and rapid progression of intima media thickness. *Arthritis Res Ther* 2010, 12(4): R158.
160. Qadri, F., Hauser, W., Jöhren, O., Dominiak, P. Kinin B₁ and B₂ receptor mRNA expression in the hypothalamus of spontaneously hypertensive rats. *Can J Physiol Pharmacol* 2002, 80(4): 258-63.

161. Couture, R., Lindsey, C.J. *Brain kallikrein-kinin system: From receptors to neuronal pathways and physiological functions*. In: Handbook of Chemical Neuroanatomy, Vol. 16, Ch. VII. Peptide Receptors. Quirion, R., Björklund, A., Hökfelt, T. (Eds.). Elsevier Science, 2000, 241-300.
162. Emanuelli, C., Chao, J., Regoli, D., Chao, L., Ni, A., Madeddu, P. *The bradykinin B1 receptor and the central regulation of blood pressure in spontaneously hypertensive rats*. Br J Pharmacol 1999, 126(8): 1769-76.
163. Alvarez, A.L., Delorenzi, A., Santajuliana, D., Finkelman, S., Nahmod, V.E., Pirola, C.J. *Central bradykinergic system in normotensive and hypertensive rats*. Clin Sci (Lond) 1992, 82(5): 513-9.
164. Campos, M.M., Cabrini, D.A., Cardozo, A.H., Rae, G.A., Toro, J.H., Calixto, J.B. *Changes in paw oedema triggered via bradykinin B(1) and B(2) receptors in streptozotocin-diabetic rats*. Eur J Pharmacol 2001, 416(1-2): 169-77.
165. Simard, B., Gabra, B.H., Sirois, P. *Inhibitory effect of a novel bradykinin B1 receptor antagonist, R-954, on enhanced vascular permeability in type 1 diabetic mice*. Can J Physiol Pharmacol 2002, 80(12): 1203-7.
166. Abdouh, M., Talbot, S., Couture, R., Hassessian, H.M. *Retinal plasma extravasation in streptozotocin-diabetic rats mediated by kinin B(1) and B(2) receptors*. Br J Pharmacol 2008, 154(1): 136-43.
167. Nadar, R., Derrick, A., Naidoo, S., Naidoo, Y., Hess, F., Bhoola, K. *Immunoreactive B1 receptors in human transbronchial tissue*. Immunopharmacology 1996, 33(1-3): 317-20.
168. Hara, D.B., Leite, D.F., Fernandes, E.S. et al. *The relevance of kinin B1 receptor upregulation in a mouse model of colitis*. Br J Pharmacol 2008, 154(6): 1276-86.
169. Marceau, F., Regoli, D. *Therapeutic options in inflammatory bowel disease: Experimental evidence of a beneficial effect of kinin B1 receptor blockade*. Br J Pharmacol 2008, 154(6): 1163-5.
170. Bonica, J.J. *The need of a taxonomy*. Pain 1979, 6(3): 247-8.
171. Julius, D., Basbaum, A.I. *Molecular mechanisms of nociception*. Nature 2001, 413(6852): 203-10.
172. Babenko, V., V., Graven-Nielsen, T., Svensson, P., Drewes, A.M., Jensen, T.S., Arendt-Nielsen, L. *Experimental human muscle pain induced by intramuscular injections of bradykinin, serotonin, and substance P*. Eur J Pain 1999, 3(2): 93-102.
173. Eisenbarth, H., Rukwied, R., Petersen, M., Schmelz, M. *Sensitization to bradykinin B1 and B2 receptor activation in UV-B irradiated human skin*. Pain 2004, 110(1-2): 197-204.
174. Ferreira, J., Beirith, A., Mori, M.A., Araujo, R.C., Bader, M., Pesquero, J.B., Calixto, J.B. *Reduced nerve injury-induced neuropathic pain in kinin B1 receptor knock-out mice*. J Neurosci 2005, 25(9): 2405-12.
175. Davis, A.J., Perkins, M.N. *The involvement of bradykinin B1 and B2 receptor mechanisms in cytokine-induced mechanical hyperalgesia in the rat*. Br J Pharmacol 1994, 113(1): 63-8.
176. Ferreira, J., Campos, M.M., Araujo, R., Bader, M., Pesquero, J.B., Calixto, J.B. *The use of kinin B1 and B2 receptor knockout mice and selective antagonists to characterize the nociceptive responses caused by kinins at the spinal level*. Neuropharmacology 2002, 43(7): 1188-97.
177. Fox, A., Wotherspoon, G., McNair, K., Hudson, L., Patel, S., Gentry, C., Winter, J. *Regulation and function of spinal and peripheral neuronal B1 bradykinin receptors in inflammatory mechanical hyperalgesia*. Pain 2003, 104(3): 683-91.
178. Abram, S.E., Yi, J., Fuchs, A., Hogan, Q.H. *Permeability of injured and intact peripheral nerves and dorsal root ganglia*. Anesthesiology 2006, 105(1): 146-53.
179. Hunskaar, S., Fasmer, O.B., Hole, K. *Formalin test in mice, a useful technique for evaluating mild analgesics*. J Neurosci Methods 1985, 14(1): 69-76.
180. Correa, C.R., Calixto, J.B. *Evidence for participation of B1 and B2 kinin receptors in formalin-induced nociceptive response in the mouse*. Br J Pharmacol 1993, 110(1): 193-8.
181. Sufka, K.J., Roach, J.T. *Stimulus properties and antinociceptive effects of selective bradykinin B1 and B2 receptor antagonists in rats*. Pain 1996, 66(1): 99-103.
182. Perkins, M.N., Campbell, E., Dray, A. *Antinociceptive activity of the bradykinin B1 and B2 receptor antagonists, des-Arg9, [Leu8]-BK and HOE 140, in two models of persistent hyperalgesia in the rat*. Pain 1993, 53(2): 191-7.
183. Perkins, M.N., Kelly, D. *Induction of bradykinin B1 receptors in vivo in a model of ultra-violet irradiation-induced thermal hyperalgesia in the rat*. Br J Pharmacol 1993, 110(4): 1441-4.
184. Ferreira, J., Campos, M.M., Pesquero, J.B., Araujo, R.C., Bader, M., Calixto, J.B. *Evidence for the participation of kinins in Freund's adjuvant-induced inflammatory and nociceptive responses in kinin B1 and B2 receptor knockout mice*. Neuropharmacology 2001, 41(8): 1006-12.
185. Cruwys, S.C., Garrett, N.E., Perkins, M.N., Blake, D.R., Kidd, B.L. *The role of bradykinin B1 receptors in the maintenance of intra-articular plasma extravasation in chronic antigen-induced arthritis*. Br J Pharmacol 1994, 113(3): 940-4.
186. Dong, H., Sun, H., Magal, E. et al. *Inflammatory pain in the rabbit: A new, efficient method for measuring mechanical hyperalgesia in the hind paw*. J Neurosci Methods 2008, 168(1): 76-87.
187. Couture, R., Harrison, M., Vianna, R.M., Cloutier, F. *Kinin receptors in pain and inflammation*. Eur J Pharmacol 2001, 429(1-3): 161-76.
188. Davis, A.J., Perkins, M.N. *Induction of B1 receptors in vivo in a model of persistent inflammatory mechanical hyperalgesia in the rat*. Neuropharmacology 1994, 33(1): 127-33.
189. Blais, C., Jr., Couture, R., Drapeau, G., Colman, R.W., Adam, A. *Involvement of endogenous kinins in the pathogenesis of peptidoglycan-induced arthritis in the Lewis rat*. Arthritis Rheum 1997, 40(7): 1327-33.
190. Eisen, V. *Plasma kinins in synovial exudates*. Br J Exp Pathol 1970, 51(3): 322-7.
191. Bond, A.P., Lemon, M., Dieppe, P.A., Bhoola, K.D. *Generation of kinins in synovial fluid from patients with arthropathy*. Immunopharmacology 1997, 36(2-3): 209-16.
192. Cassim, B., Shaw, O.M., Mazur, M. et al. *Kallikreins, kininogens and kinin receptors on circulating and synovial fluid neutrophils: Role in kinin generation in rheumatoid arthritis*. Rheumatology (Oxford) 2009, 48(5): 490-6.
193. Sevcik, M.A., Ghilardi, J.R., Halvorson, K.G., Lindsay, T.H., Kubota, K., Mantyh, P.W. *Analgesic efficacy of bradykinin B1 antagonists in a murine bone cancer pain model*. J Pain 2005, 6(11): 771-5.
194. Bennett, G.J., Xie, Y.K. *A peripheral mononeuropathy in rat that produces disorders of pain sensation like those seen in man*. Pain 1988, 33(1): 87-107.
195. Seltzer, Z., Dubner, R., Shir, Y. *A novel behavioral model of neuropathic pain disorders produced in rats by partial sciatic nerve injury*. Pain 1990, 43(2): 205-18.
196. Kim, S.H., Chung, J.M. *An experimental model for peripheral neuropathy produced by segmental spinal nerve ligation in the rat*. Pain 1992, 50(3): 355-63.
197. Levy, D., Zochodne, D.W. *Increased mRNA expression of the B1 and B2 bradykinin receptors and antinociceptive effects of their antagonists in an animal model of neuropathic pain*. Pain 2000, 86(3): 265-71.
198. Yamaguchi-Sase, S., Hayashi, I., Okamoto, H., Nara, Y., Matsuzaki, S., Hoka, S., Majima, M. *Amelioration of hyperalgesia by kinin receptor antagonists or kininogen deficiency in chronic constriction nerve injury in rats*. Inflamm Res 2003, 52(4): 164-9.

199. Quintao, N.L., Passos, G.F., Medeiros, R. et al. *Neuropathic pain-like behavior after brachial plexus avulsion in mice: The relevance of kinin B1 and B2 receptors*. J Neurosci 2008, 28(11): 2856-63.
200. Luiz, A.P., Schroeder, S.D., Chichorro, J.G., Calixto, J.B., Zampronio, A.R., Rae, G.A. *Kinin B(1) and B(2) receptors contribute to orofacial heat hyperalgesia induced by infraorbital nerve constriction injury in mice and rats*. Neuropeptides 2010, 44(2): 87-92.
201. Campos, M.M., Ongali, B., De Souza, B.H., Schanstra, J.P., Girolami, J.P., Chabot, J.G., Couture, R. *Expression and distribution of kinin B1 receptor in the rat brain and alterations induced by diabetes in the model of streptozotocin*. Synapse 2005, 57(1): 29-37.
202. Catanzaro, O.L., Dziubecki, D., Labal, E., Sirois, P. *Activation of peritoneal macrophages during the evolution of type 1 diabetes (insulinitis) in streptozotocin-treated mice*. Peptides 2010, 31(10): 1884-7.
203. Cloutier, F., Couture, R. *Pharmacological characterization of the cardiovascular responses elicited by kinin B(1) and B(2) receptor agonists in the spinal cord of streptozotocin-diabetic rats*. Br J Pharmacol 2000, 130(2): 375-85.
204. Gabra, B.H., Sirois, P. *Role of bradykinin B(1) receptors in diabetes-induced hyperalgesia in streptozotocin-treated mice*. Eur J Pharmacol 2002, 457(2-3): 115-24.
205. Ongali, B., Campos, M.M., Petcu, M. et al. *Expression of kinin B1 receptors in the spinal cord of streptozotocin-diabetic rat*. Neuroreport 2004, 15(16): 2463-6.
206. Mage, M., Pecher, C., Neau, E. et al. *Induction of B1 receptors in streptozotocin diabetic rats: Possible involvement in the control of hyperglycemia-induced glomerular Erk 1 and 2 phosphorylation*. Can J Physiol Pharmacol 2002, 80(4): 328-33.
207. Vianna, R.M., Ongali, B., Regoli, D., Calixto, J.B., Couture, R. *Up-regulation of kinin B1 receptor in the lung of streptozotocin-diabetic rat: Autoradiographic and functional evidence*. Br J Pharmacol 2003, 138(1): 13-22.
208. Dias, J.P., Ismael, M.A., Pilon, M., de Champlain, J., Ferrari, B., Carayon, P., Couture, R. *The kinin B1 receptor antagonist SSR240612 reverses tactile and cold allodynia in an experimental rat model of insulin resistance*. Br J Pharmacol 2007, 152(2): 280-7.
209. Talbot, S., Chahmi, E., Dias, J.P., Couture, R. *Key role for spinal dorsal horn microglial kinin B1 receptor in early diabetic pain neuropathy*. J Neuroinflammation 2010, 7(1): 36.
210. Gabra, B.H., Berthiaume, N., Sirois, P., Nantel, F., Battistini, B. *The kinin system mediates hyperalgesia through the inducible bradykinin B1 receptor subtype: Evidence in various experimental animal models of type 1 and type 2 diabetic neuropathy*. Biol Chem 2006, 387(2): 127-43.
211. Gabra, B.H., Sirois, P. *Hyperalgesia in non-obese diabetic (NOD) mice: A role for the inducible bradykinin B1 receptor*. Eur J Pharmacol 2005, 514(1): 61-7.
212. Gabra, B.H., Sirois, P. *Beneficial effect of chronic treatment with the selective bradykinin B1 receptor antagonists, R-715 and R-954, in attenuating streptozotocin-diabetic thermal hyperalgesia in mice*. Peptides 2003, 24(8): 1131-9.
213. Dias, J.P., Talbot, S., Senecal, J., Carayon, P., Couture, R. *Kinin B1 receptor enhances the oxidative stress in a rat model of insulin resistance: Outcome in hypertension, allodynia and metabolic complications*. PLoS ONE 2010, 5(9): e12622.
214. Gabra, B.H., Merino, V.F., Bader, M., Pesquero, J.B., Sirois, P. *Absence of diabetic hyperalgesia in bradykinin B1 receptor-knockout mice*. Regul Pept 2005, 127(1-3): 245-8.
215. Chen, J.J., Johnson, E.J. *Targeting the bradykinin B1 receptor to reduce pain*. Expert Opin Ther Targets 2007, 11(1): 21-35.
216. Fincham, C.I., Bressan, A., Paris, M., Rossi, C., Fattori, D. *Bradykinin receptor antagonists—A review of the patent literature 2005-2008*. Expert Opin Ther Pat 2009, 19(7): 919-41.
217. Ferrari, B., Gougat, J., Muneaux, C., Muneaux, Y., Perreaut, P., Planchenault, C. (sanofi-aventis). *Novel N-(arylsulphonyl)amino acid derivatives having bradykinin receptor affinity*. WO 1997025315.
218. Ferrari, B., Gougat, J., Muneaux, Y., Perreaut, P., Sarran, L. (sanofi-aventis). *Derivatives of N-(arylsulfonyl)beta-aminoacids comprising a substituted aminomethyl group*. WO 2002076964.
219. Kuduk, S.D., Bock, M.G. *Bradykinin B1 receptor antagonists as novel analgesics: A retrospective of selected medicinal chemistry developments*. Curr Top Med Chem 2008, 8(16): 1420-30.
220. Tang, C., Carr, B.A., Poignant, F. et al. *CYP2C75-involved autoinduction of metabolism in rhesus monkeys of methyl 3-chloro-3'-fluoro-4'-{[(1R)-1-[[[1-(trifluoroacetyl)amino]cyclopropyl]carbonyl]amino]ethyl}-1,1'-biphenyl-2-carboxylate (MK-0686), a bradykinin B1 receptor antagonist*. J Pharmacol Exp Ther 2008, 325(3): 935-46.
221. Bodmer-Narkevitch, V., Anthony, N.J., Cofre, V. et al. *Indazole derivatives as novel bradykinin B(1) receptor antagonists*. Bioorg Med Chem Lett 2010, 20(23): 7011-4.
222. Feng, D.M., DiPardo, R.M., Wai, J.M. et al. *A new class of bradykinin B1 receptor antagonists with high oral bioavailability and minimal PXR activity*. Bioorg Med Chem Lett 2008, 18(2): 682-7.
223. Kuduk, S.D., Chang, R.K., DiPardo, R.M. et al. *Bradykinin B1 receptor antagonists: An alpha-hydroxy amide with an improved metabolism profile*. Bioorg Med Chem Lett 2008, 18(18): 5107-10.
224. Wood, M.R., Schirripa, K.M., Kim, J.J. et al. *Alpha-hydroxy amides as a novel class of bradykinin B1 selective antagonists*. Bioorg Med Chem Lett 2008, 18(2): 716-20.
225. Li, G., Hodgetts, K., Chenard, B.L. (Neurogen Corp.). *Aryl sulfones*. WO 2008002849.
226. Schaudt, M., Locardi, E., Zischinsky, G. et al. *Novel small molecule bradykinin B1 receptor antagonists. Part 1: Benzamides and semicarbazides*. Bioorg Med Chem Lett 2010, 20(3): 1225-8.